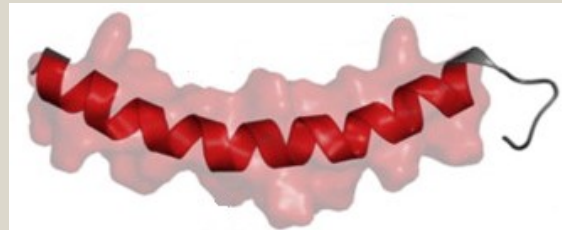


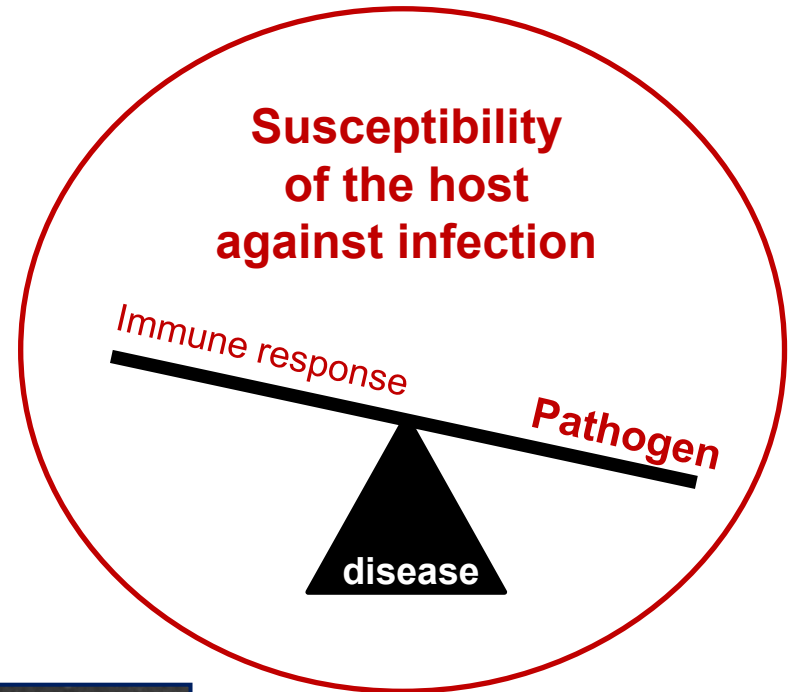
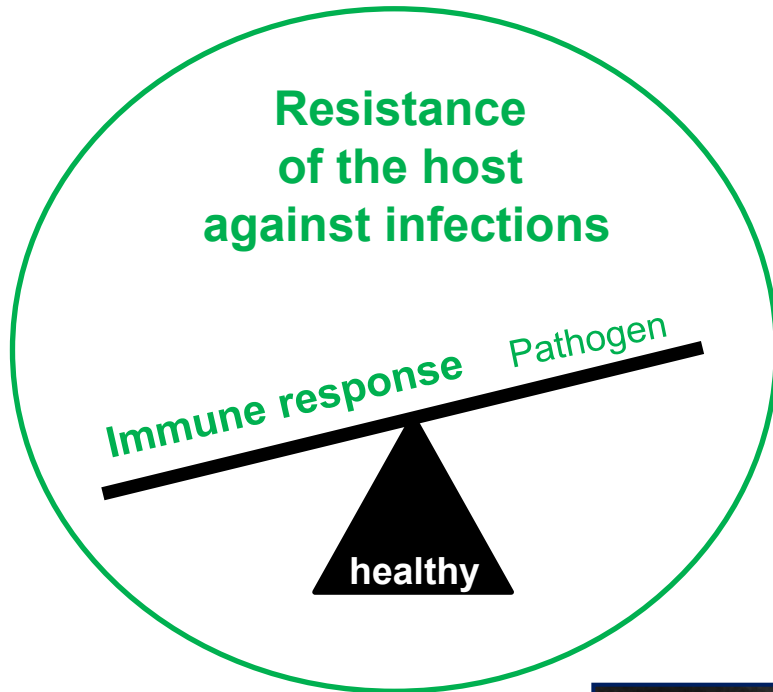
# Wie alte Waffen helfen, neue therapeutische Ansätze gegen Infektionskrankheiten zu finden

*„Old weapons against infectious diseases“*

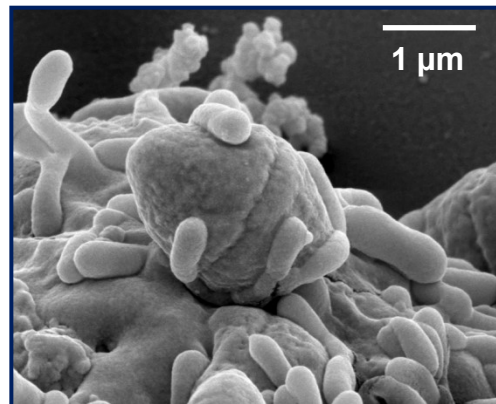


Prof. Dr. Maren von Köckritz-Blickwede,  
Infection Biochemistry,  
Department of Biochemistry,  
TiHo Hannover, Germany

# Infections in human and animals



*Streptococcus pyogenes* entrapped by immune cell



von Köckritz-Blickwede et al., *Blood*, 2008

# Alternatives

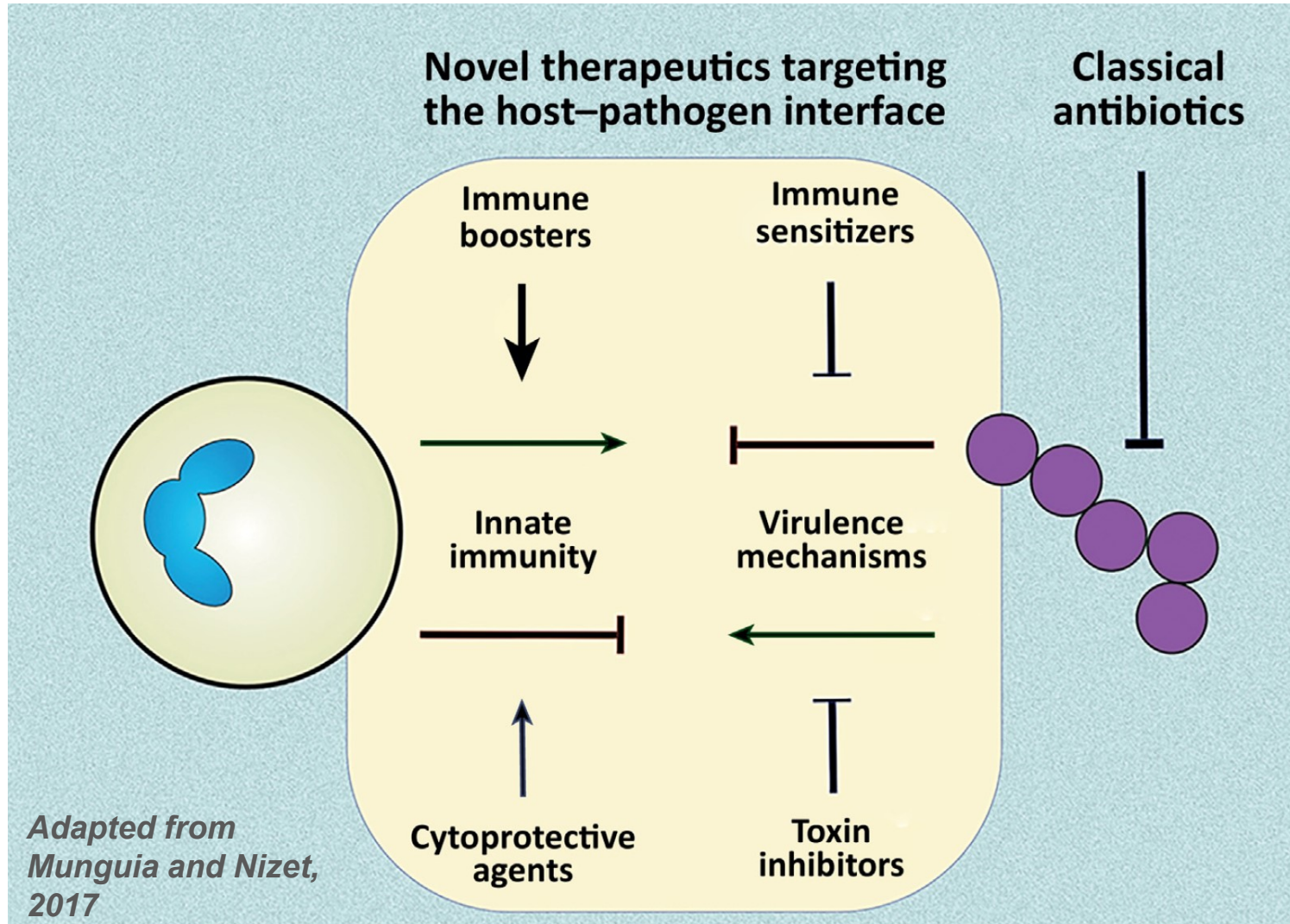
---

Development of drugs which  
**boost the host immune system**  
used alongside conventional antibiotic treatment  
for successful therapy of difficult infections.



[http://www.loadmedical.com/  
media/landingpages/Immun/  
Immun-00005.jpg](http://www.loadmedical.com/media/landingpages/Immun/Immun-00005.jpg)

# Alternatives



Adapted from  
Munguia and Nizet,  
2017

# Guardian cell of the immune system

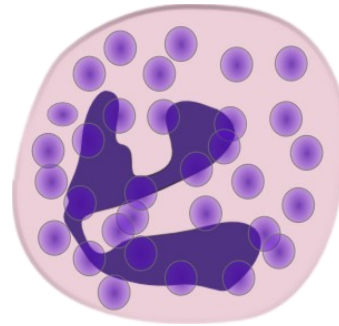
## Neutrophil

### Phagocytosis

(intracellular uptake and killing)

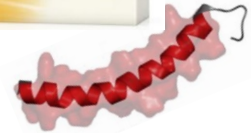


Guarea kunthiana



### Degranulation

(extracellular release of antimicrobial peptides and others)



### Neutrophil extracellular traps (NETs)

(extracellular entrapment and killing of bacteria)



# Phagocytosis of bacteria

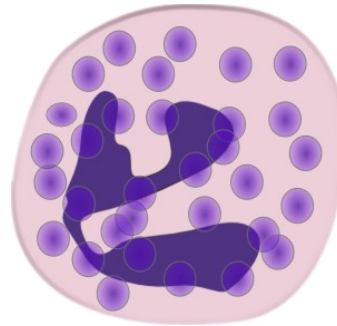
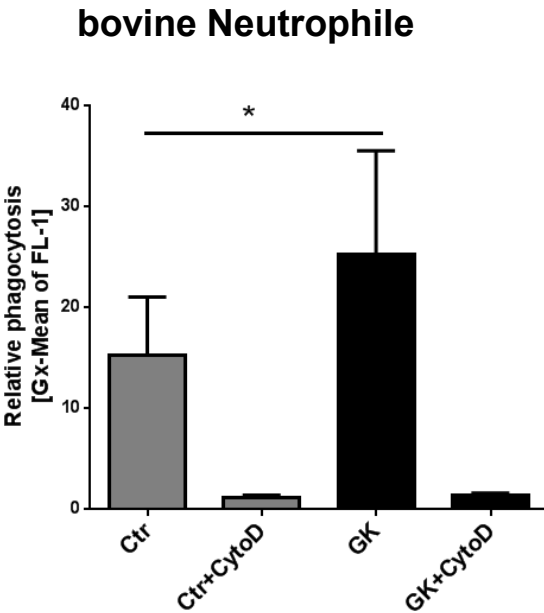
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## PHAGOCYTOSIS

To defend the body against bacteria, human neutrophils (white blood cells) ingest and kill invading pathogens like this *E. coli*

# Natural products that stimulate phagocytosis

## Neutrophil



CytoD: blocks phagocytosis  
GK: Guarea kunthiana extracts

## Defense against pathogens

*Jerjomiceva, 2014*

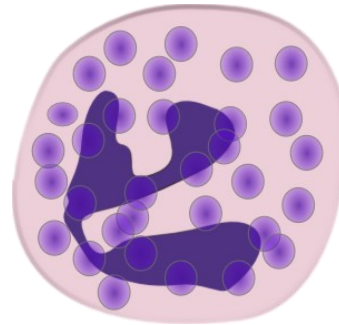
# Guardian cell of the immune system

## Neutrophil

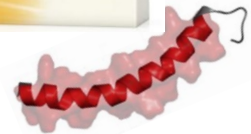
**Phagocytosis**  
(intracellular uptake and killing)



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**Degranulation**  
(extracellular release of antimicrobial peptides and others)



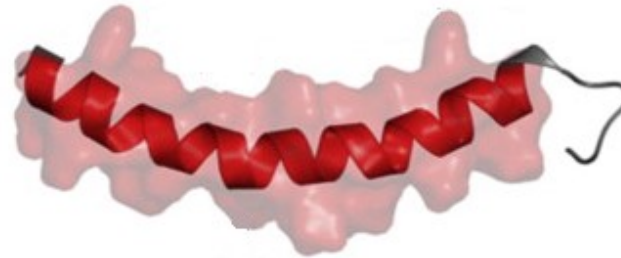
**Neutrophil extracellular traps (NETs)**  
(extracellular entrapment and killing of bacteria)



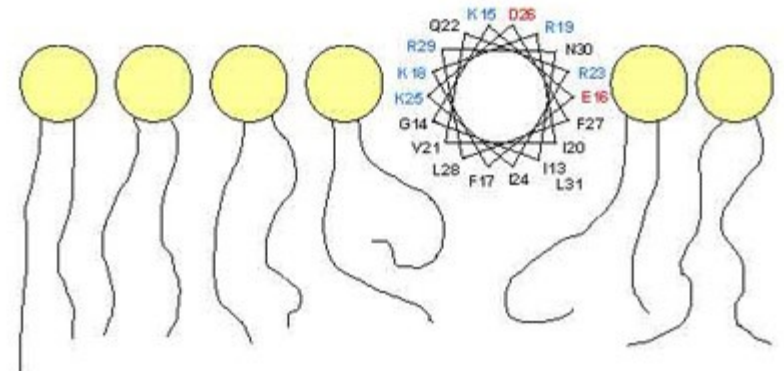
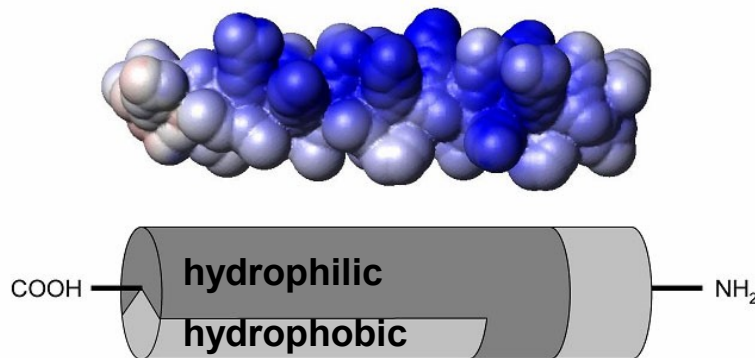


# Host antimicrobial peptides (AMPs)

- Small molecules, positively charged, amphipathic structure
- Released by neutrophils and other cells e.g. keratinocytes
- e.g. cathelicidin LL-37

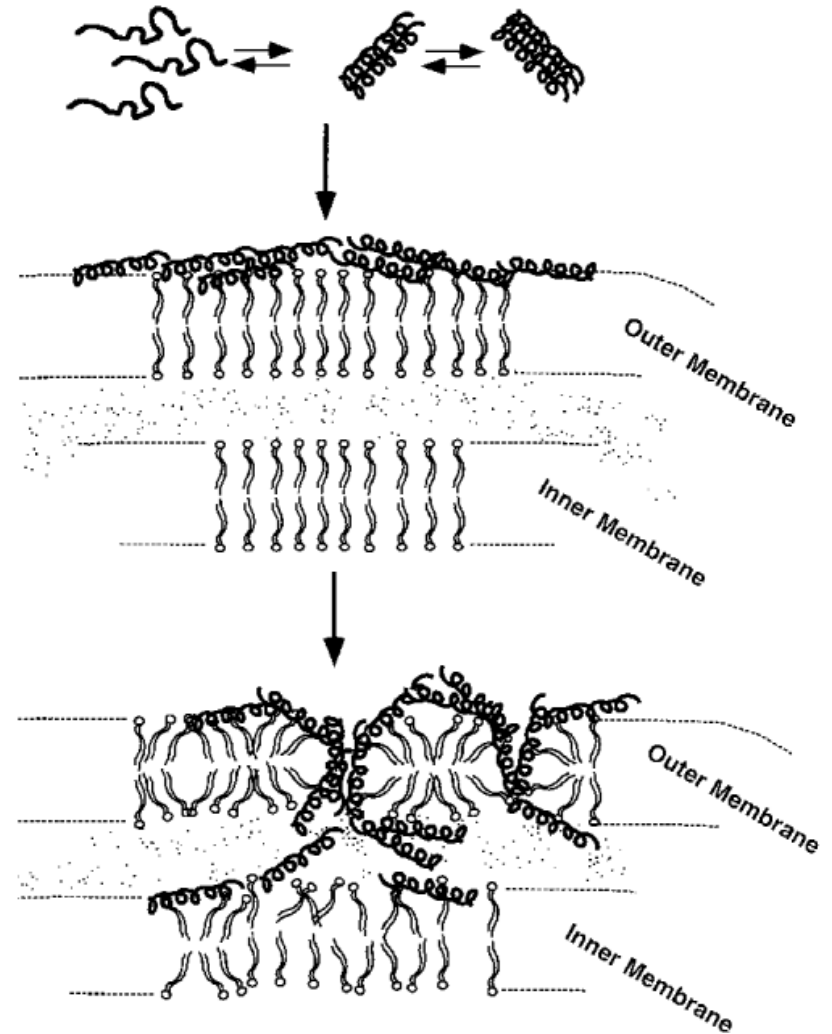
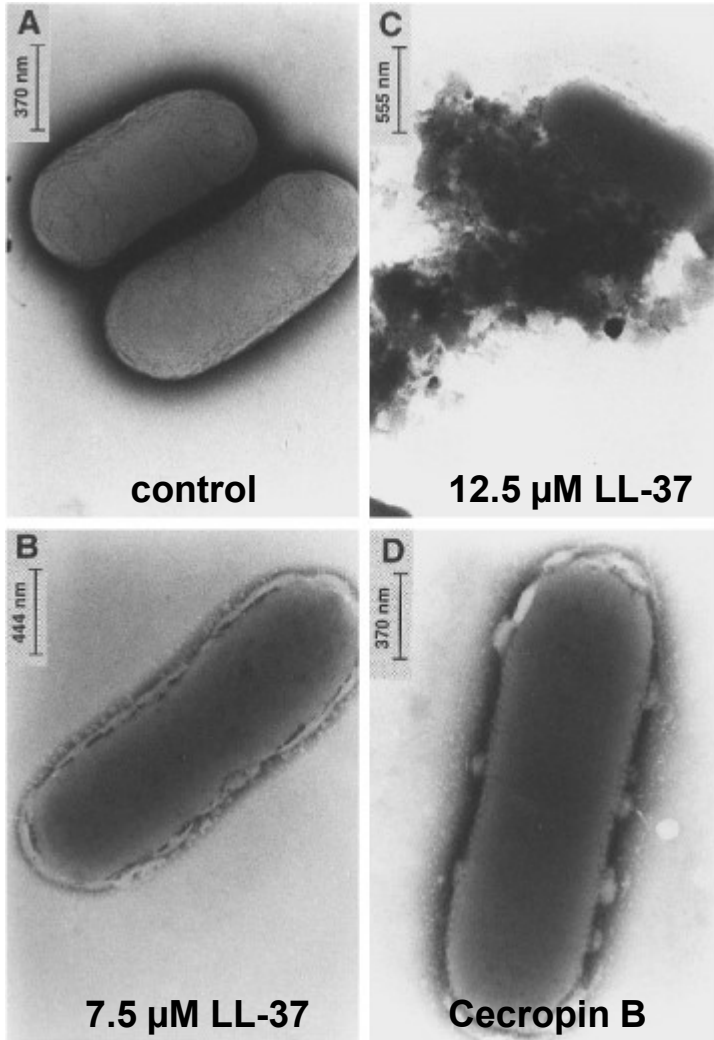


*Vandamme et al., 2012*



*Henzler Wildman, 2003*

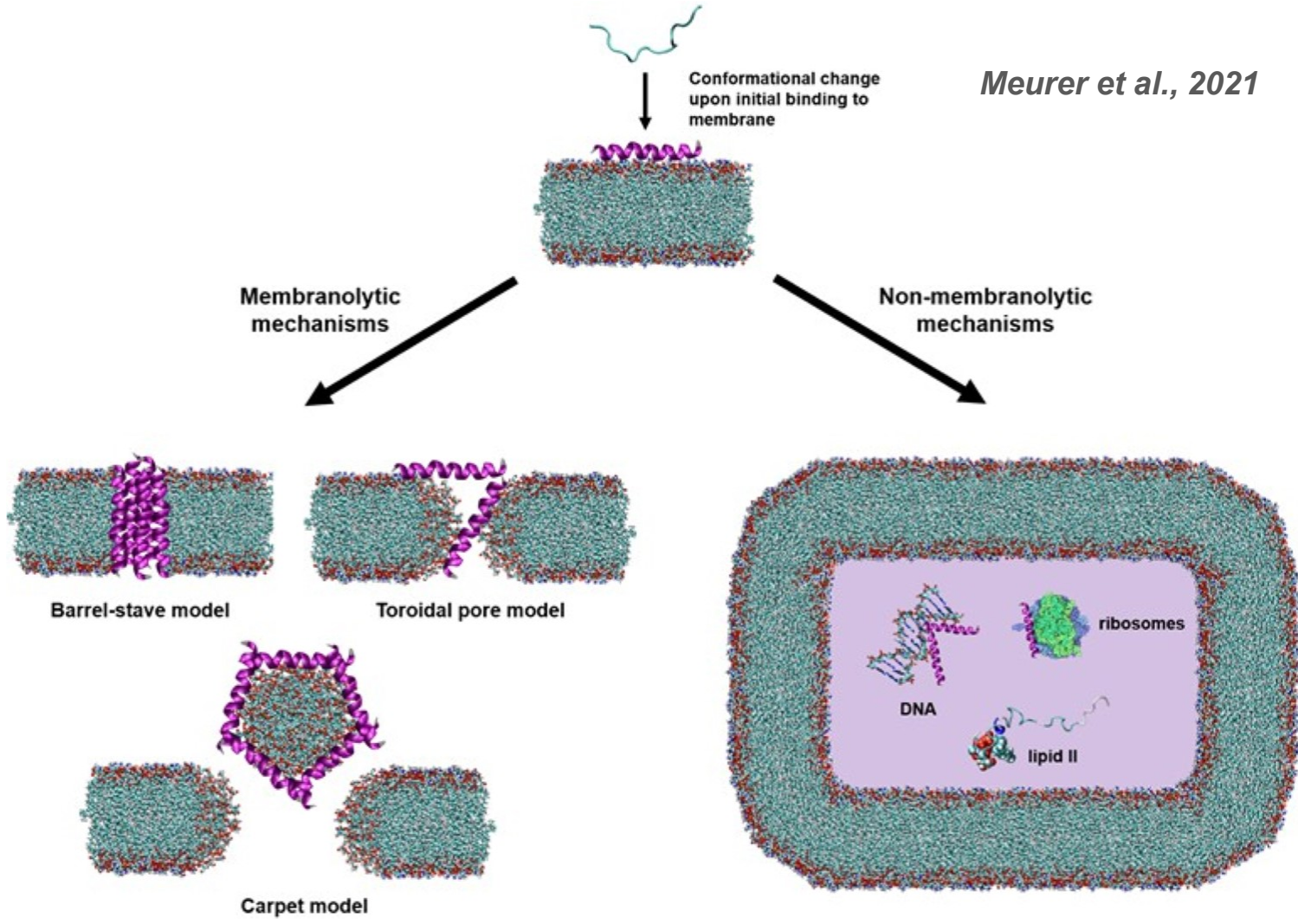
# Host antimicrobial peptides (AMPs)



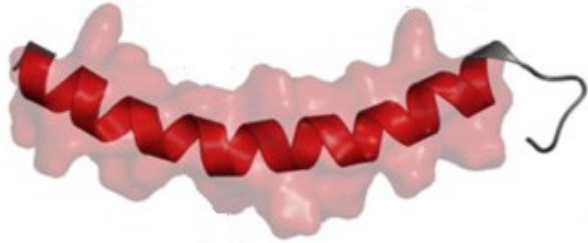
Oren, 1999

# Host antimicrobial peptides (AMPs)

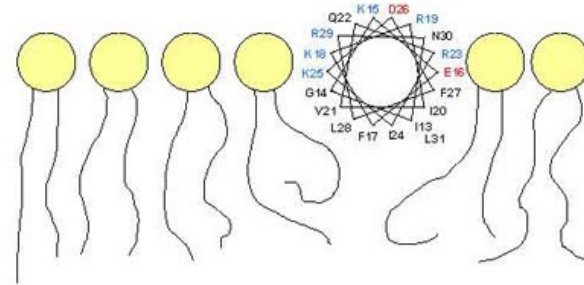
*Meurer et al., 2021*



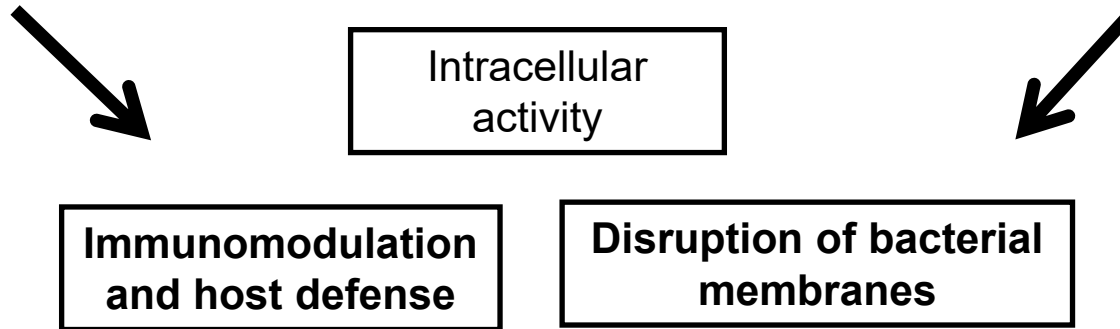
# Antimicrobial peptides



Vandamme et al., 2012



Henzler Wildman, 2003



## Advantage of AMPs:

## Low propensity to select resistance



# The effect of antimicrobial peptides (AMPs) on the innate immune response against zoonotic *S. aureus*

Stefanie Blodkamp

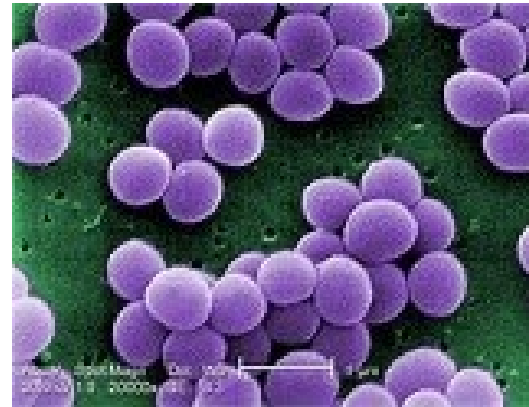
FRIEDRICH-LOEFFLER-INSTITUT

**FLI**

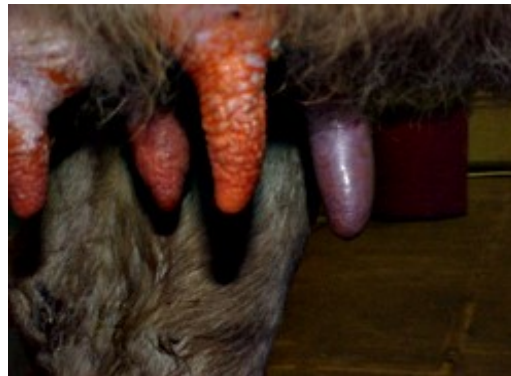
Bundesforschungsinstitut für Tiergesundheit  
Federal Research Institute for Animal Health

# *Staphylococcus aureus*

- Round-shaped, facultative anaerobic
- Typical commensal
- Skin & soft tissue infections
- Necrotizing pneumonia
- Mastitis



[https://en.wikipedia.org/wiki/Staphylococcus\\_aureus](https://en.wikipedia.org/wiki/Staphylococcus_aureus)



<http://cal.vet.upenn.edu/projects/fieldservice/Dairy/Mastitis/staphaur.htm>



[http://www.20min.ch/diashow/95650/95650wg7Hlrp\\_Q2zu7SJS4NegBw.jpg](http://www.20min.ch/diashow/95650/95650wg7Hlrp_Q2zu7SJS4NegBw.jpg)

# Methicillin-resistant *Staphylococcus aureus* (MRSA)

HA-MRSA = healthcare-associated MRSA

CA-MRSA = community-associated MRSA

LA-MRSA = livestock-associated MRSA



# Methicillin-resistant *Staphylococcus aureus* (MRSA)

## Goal:





1. AMP susceptibility testing of LA-MRSA field isolates
2. Factors that alter AMP MICs of *S. aureus*

LA-MRSA = livestock-associated MRSA










# Cathelicidins

Host	AMP	Sequence
	LL-37	LLGDFFRKSKEKIGKEFKRIVQRIKDFLRNLPRTES
	mCRAMP	GLLRKGGGEKIGEKLLKKIGQKIKNFFQKLVPQPE
	CAP18	GLRKRLRKFRNKIKEKLLKKIGQKIQGFVPKLPRTDY
	BMAP-27	GRFKRFRKKFKKLFKKLSPVIPLLHLG
	BMAP-28	GGLRSLGRKILRAWKKYGPIIVPIIRIG

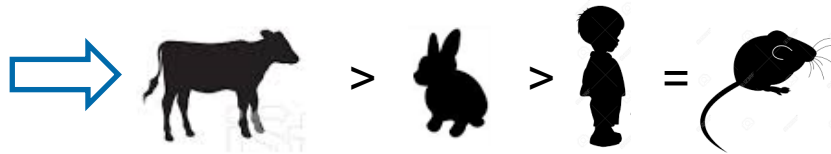
# AMP MICs of field isolates





Source	No. of isolates	Mode MIC [mg/L]				
		LL-37 	mCRAMP 	CAP18 	BMAP-27 	BMAP-28 
Cattle d.	16	≥128	≥128	32	8	2
Chicken h.	19	≥128	≥128	64	8	2
Chicken d.	5	≥128	≥128	≥128	16	8
Turkey d.	17	≥128	≥128	64	8	4, 8
Swine d.	32	≥128	≥128	64	8	2
Farm e.	31	≥128	≥128	64	8	2
Chicken meat	10	≥128	≥128	64	16	4
Turkey meat	21	≥128	≥128	64	8	4

d. = diseased; e. = environment and healthy employees; h. = healthy

# Antimicrobial peptides

- Antimicrobial activity of cathelicidins derived from different hosts



Host	AMP	Sequence	Hydrophobic residues %
	LL-37	LLGDFFRKSKEKIGKEFKRIVQRIKDFLRNLPRTES	35
	mCRAMP	GLLRKGGEKIGEKLLKIGQKKNFFQKLVPQPE	29
	CAP18	GLRKRLRKFRNKIKEKLLKIGQKIQGFVPKLAPRTDY	29
	BMAP-27	GRFKRFRKKFKKLFKKLSPVIPLLHLG	40
	BMAP-28	GGLRSLGRKILRAWKKGPIIVPIIRIG	42

*Blodkamp et al., Vet. Micro., 2015*

# Bovine cathelicidins

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## BMAP-27



*Baumann et al., 2017*



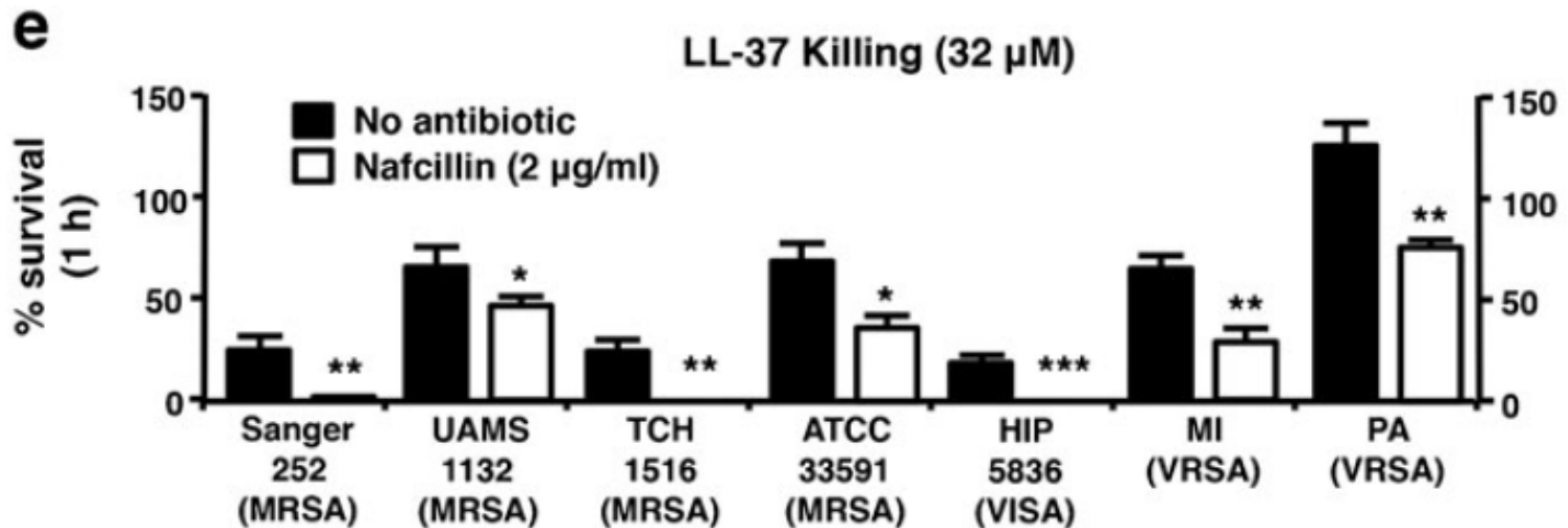
## BMAP-28

Thivierge et al., 2013



# Antimicrobial peptides

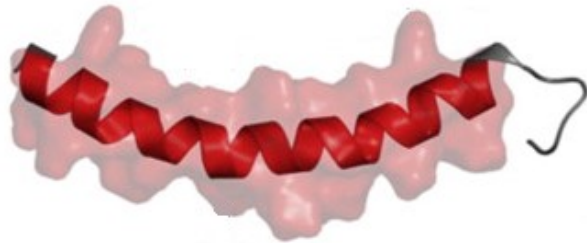
## Synergisms of AMPs with antibiotics



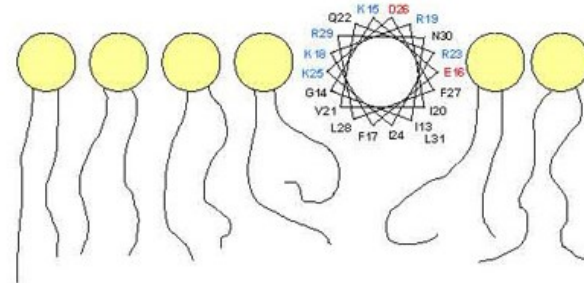
→ Nafcillin enhances killing of methicillin-resistant *Staphylococcus aureus* by cathelicidins

Sakoulas et al., 2014

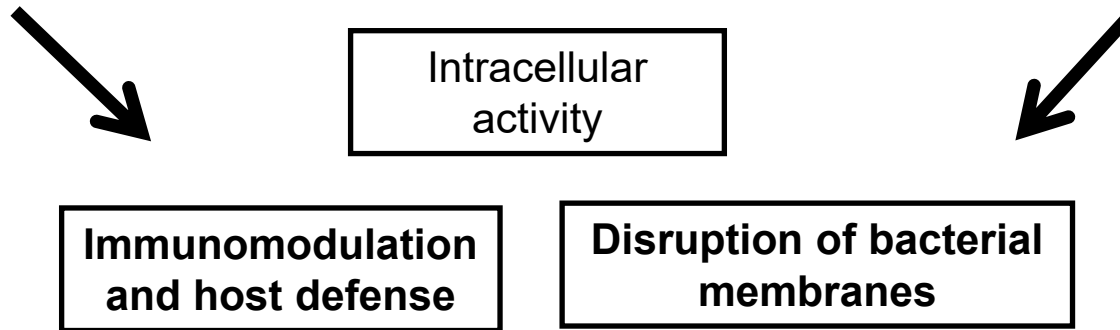
# Antimicrobial peptides



Vandamme et al., 2012



Henzler Wildman, 2003



## Advantage of AMPs:

**Synergistic effects with antibiotics  
and other immune defence mechanisms**

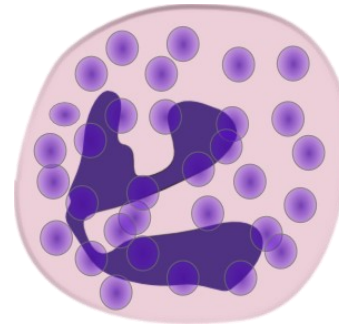
# Guardian cell of the immune system

## Neutrophil

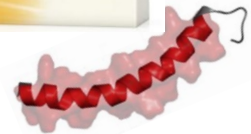
**Phagocytosis**  
(intracellular uptake and killing)



Guarea kunthiana



**Degranulation**  
(extracellular release of antimicrobial peptides and others)

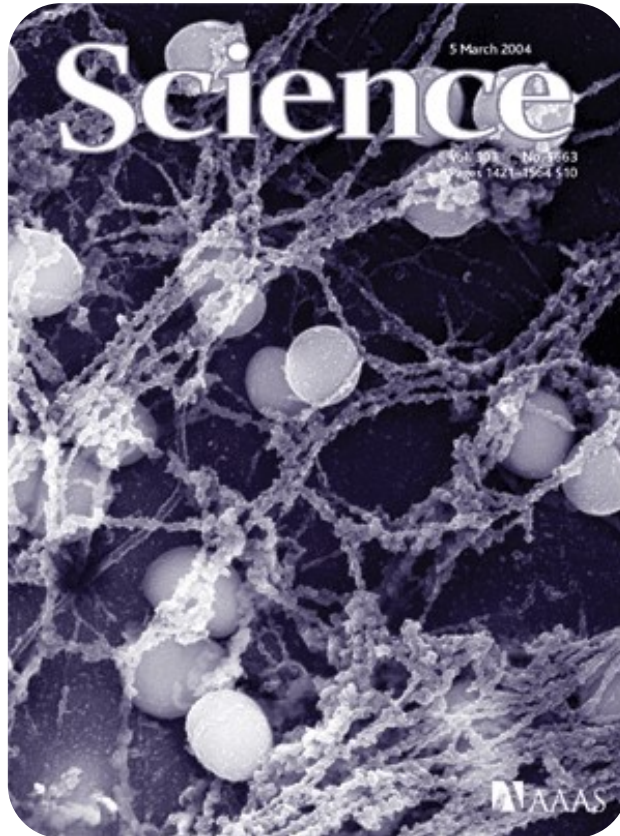


**Neutrophil extracellular traps (NETs)**  
(extracellular entrapment and killing of bacteria)



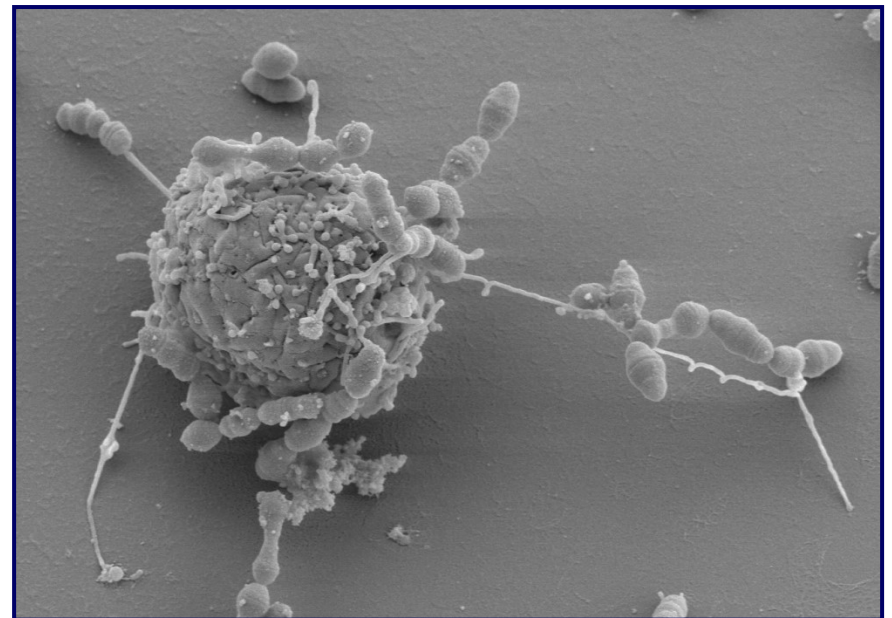
# Neutrophil extracellular traps (NETs)

## Neutrophils



*Brinkmann et al., 2004*

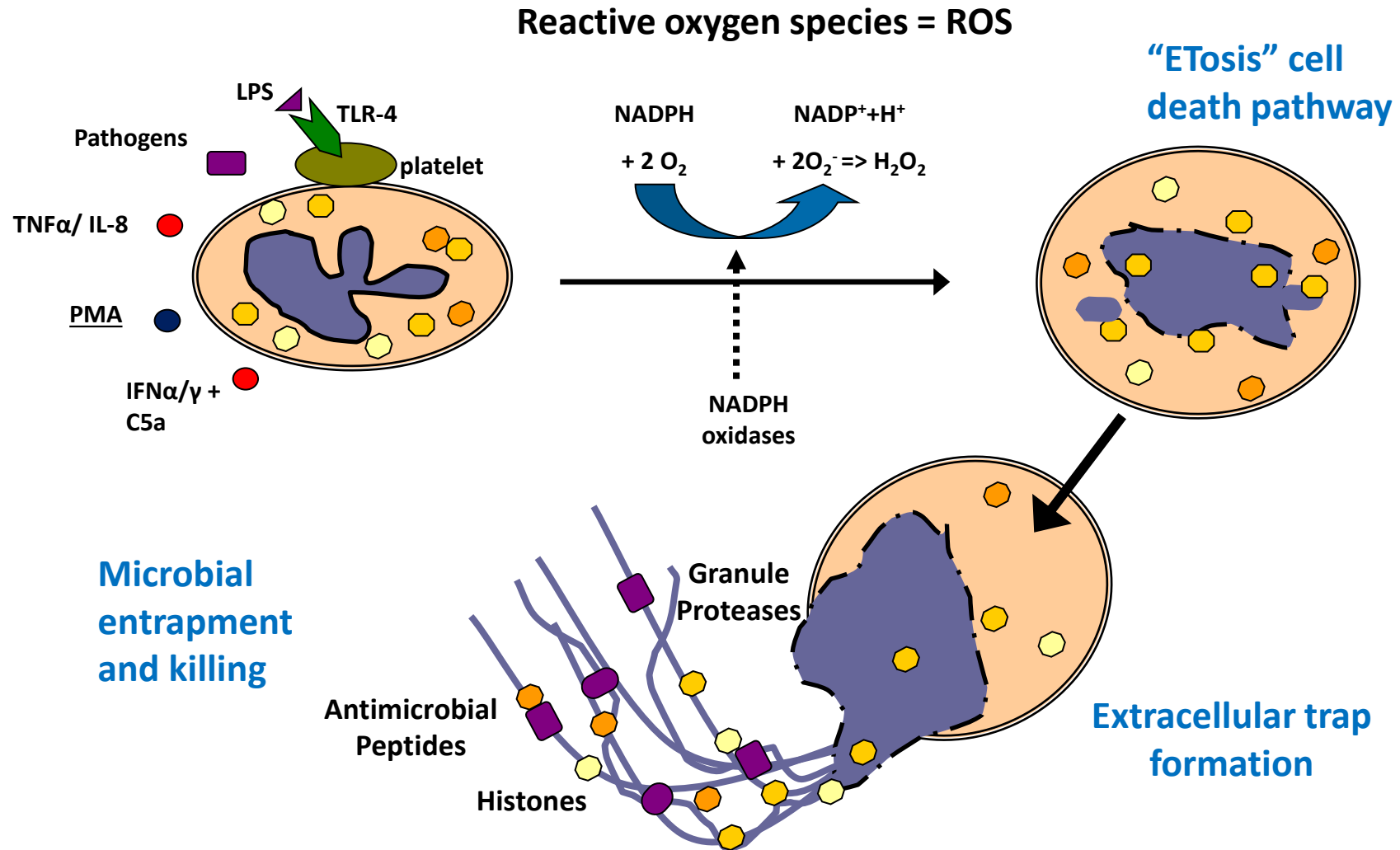
## Mast cells



*von Köckritz-Blickwede et al., Blood, 2008*



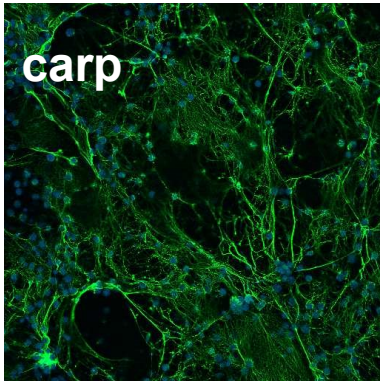
# Extracellular DNA traps (NETs)



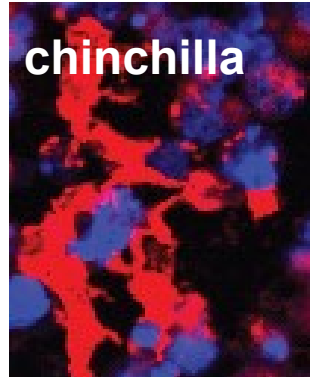
PMA = phorbol myristate acetate

*von Köckritz-Blickwede and Nizet , 2009*

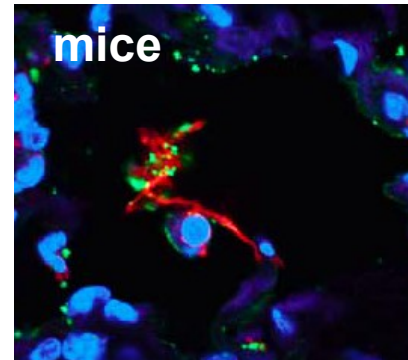
# Extracellular traps in human and animals



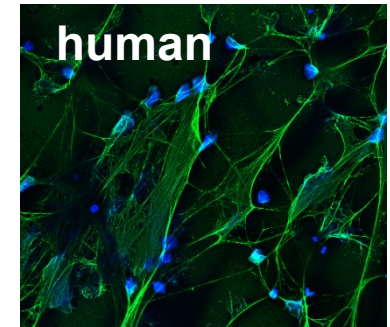
*Brogden et al., Dev. Comp. Immunol., 2014;*  
*Brogden et al., Fish and Shellfish Immunol. 2012*



*Short et al., Infect. Immun., 2013*



*Berends et al., J. Innate Immun., 2010*



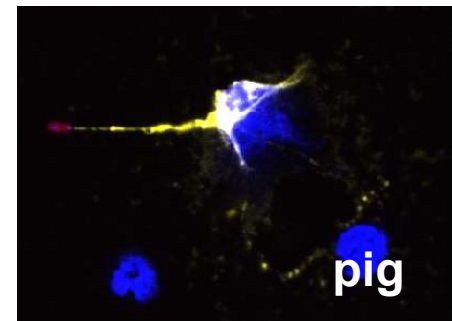
*Neumann et al., J. Innate Immun., 2014;*  
*Branitzki-Heinemann et al., Biochem. J. 2012*



*Jerjomiceva et al., J. Innate Immun., 2014*

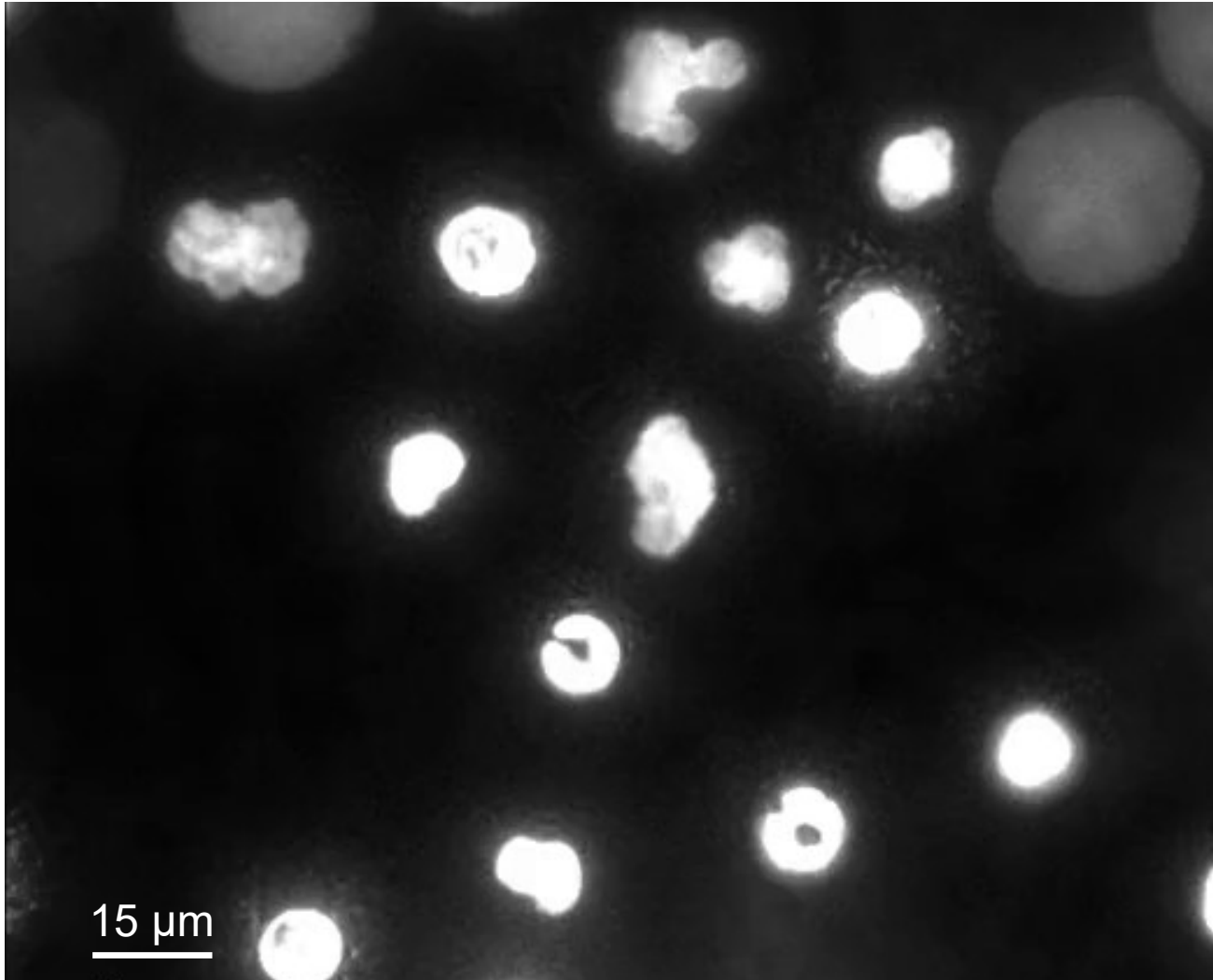


*Fingerhut et al., 2021*



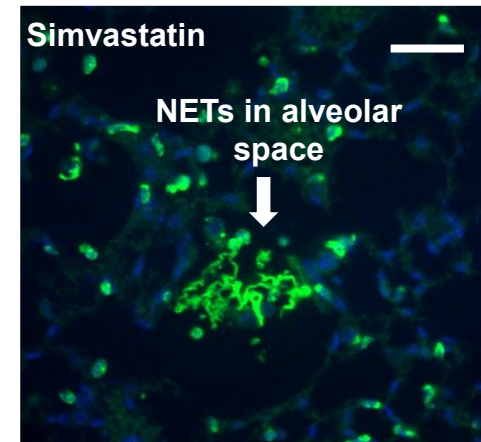
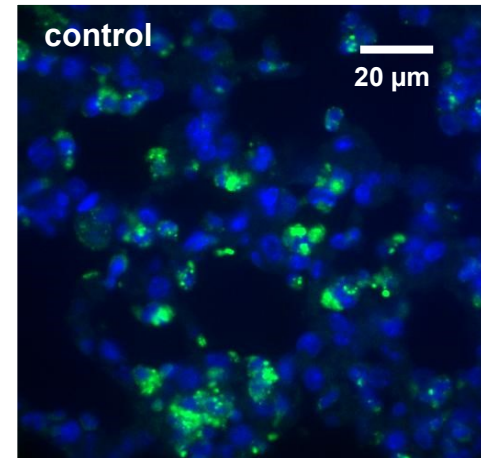
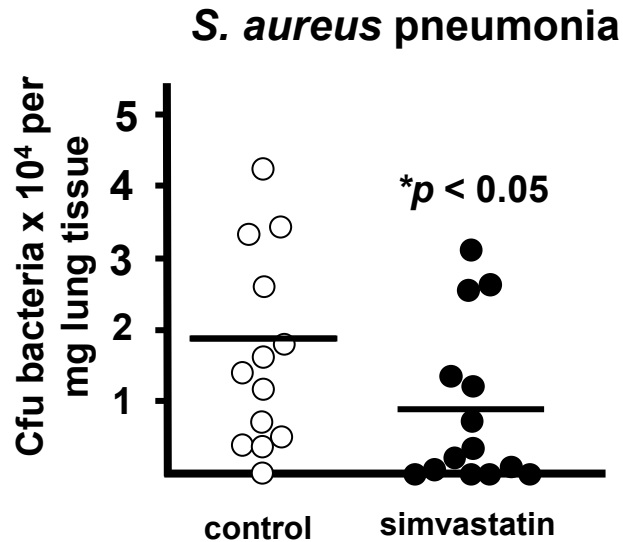
*Meurer et al., Microbiology, 2020*

# Statins induce NETs



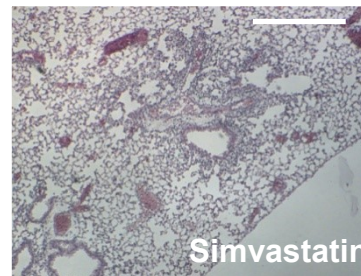
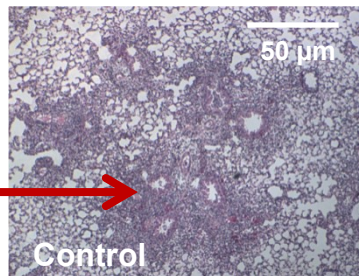
# Protective effect of NETs *in vivo*?

**Murine  
*in vivo*  
model**



**Blue: nucleus**  
**Green: antimicrobial peptide mCRAMP**

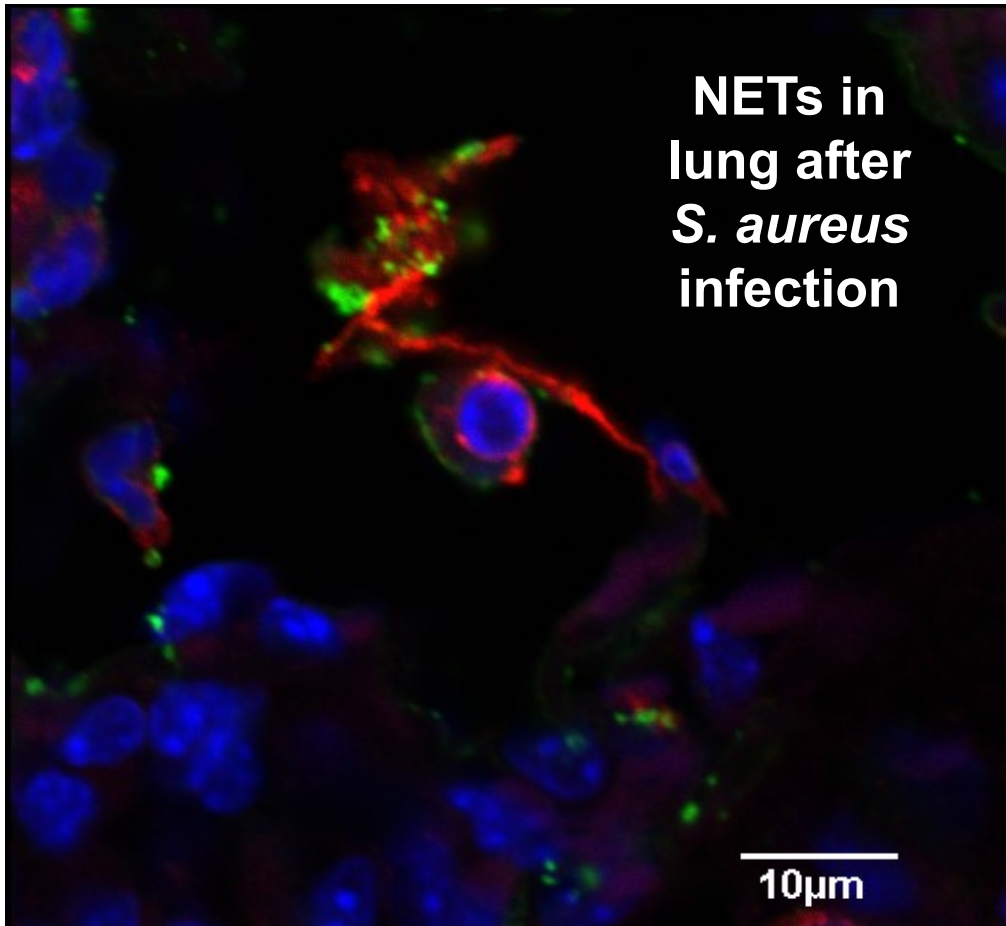
Inflammation of lung tissue



Mice were prefed for 5 days with 500 mg/kg/day statins and then infected with a sublethal dose of  $2 \times 10^8$  cfu *S. aureus* Newman intranasally for 48 h.

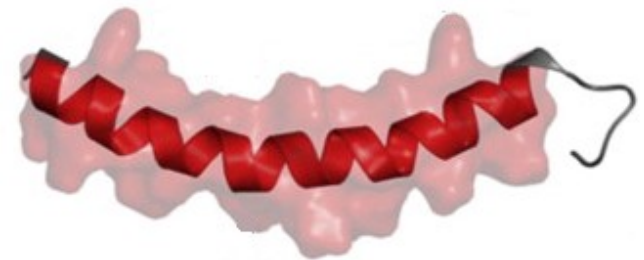
*Chow et al., 2010*

# In vivo NET-formation during infection



**Blue:** nucleus  
**Green:** histone-DNA-complex  
**Red:** antimicrobial peptide mCRAMP

**Host antimicrobial peptides (AMPs)**

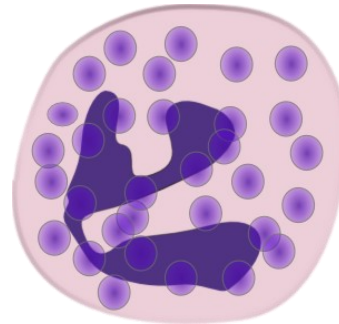
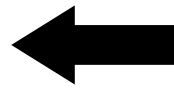


*Vandamme et al., 2012*

# Guardian cell of the immune system

## Neutrophil

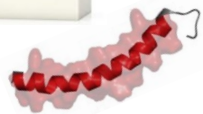
**Phagocytosis**  
(intracellular uptake and killing)



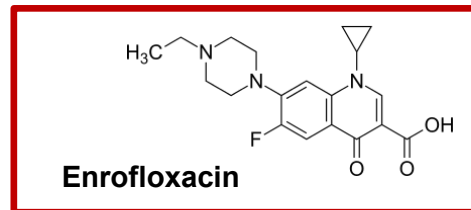
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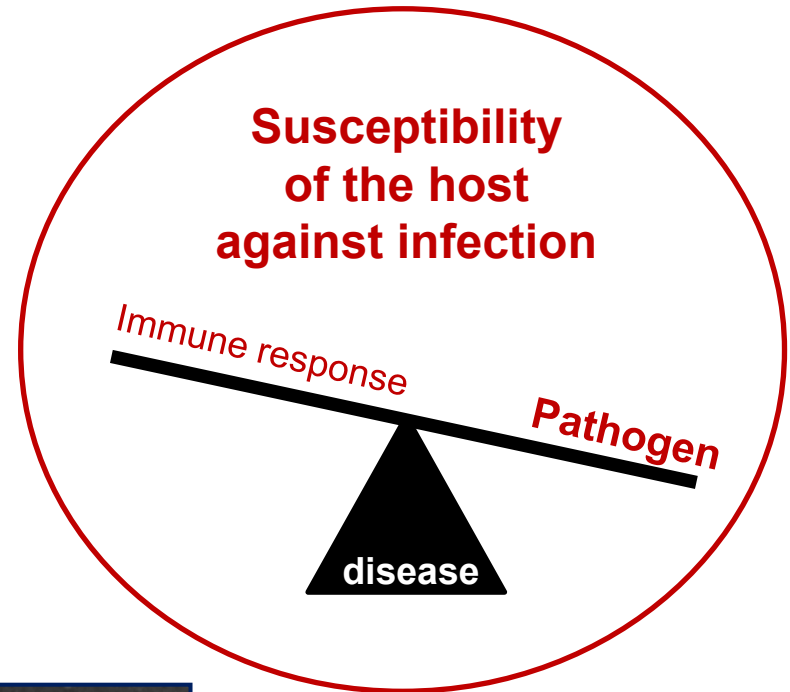
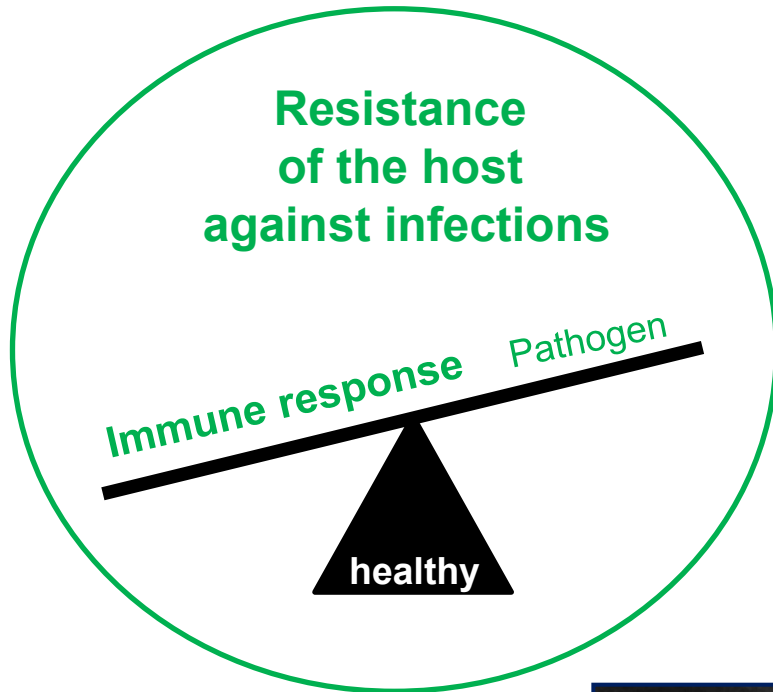
Guarea kunthiana



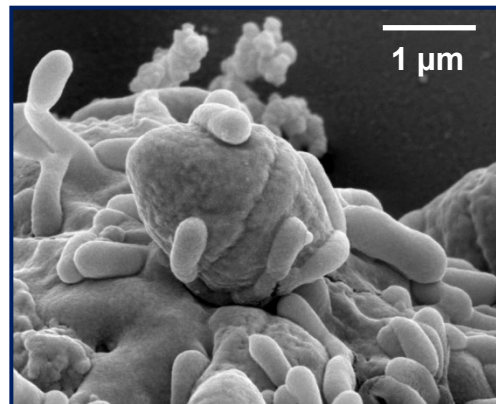
**Neutrophil extracellular traps (NETs)**  
(extracellular entrapment and killing of bacteria)



# Infections in human and animals



*Streptococcus pyogenes* entrapped by immune cell

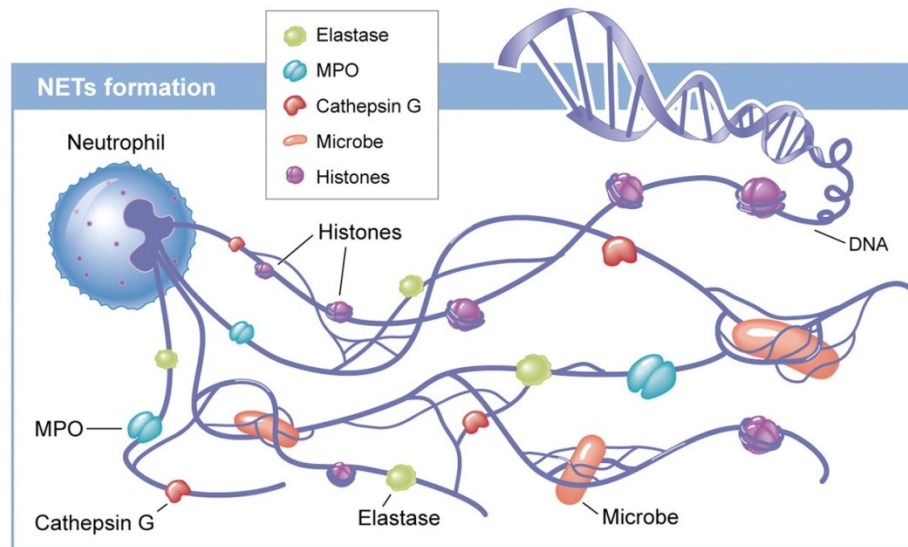


von Köckritz-Blickwede et al., *Blood*, 2008

# Balance of NETs and AMPs

## Protective defence of NETs against infections:

e.g. *Staphylococcus aureus* (Berends et al., 2010), *Streptococcus pyogenes* (Lauth et al., 2009), *Streptococcus suis* (de Buhr et al., 2014), *Yersinia enterocolitica* and *Escherichia coli* (Möllerherm and Neumann et al., 2015)



## Detrimental consequences of NETs for the host during infections with:

e.g. *Strep. pneumoniae* + influenza (Short et al., 2013)

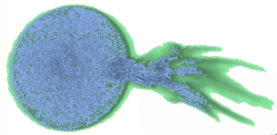
- Entrapment of pathogens
- Growth inhibition of pathogens
- Limit acute inflammation



- Cytotoxicity
- Thrombosis
- Autoimmune reactions
- Chronic inflammation



# NETs & AMPs interact with SARS-CoV-2



BRIEF DEFINITIVE REPORT

## SARS-CoV-2-triggered neutrophil extracellular traps mediate COVID-19 pathology

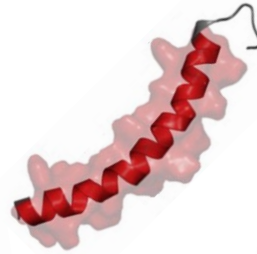
Flavio Protásio Veras<sup>1,2</sup>, Marjorie Cordeiro Pontelli<sup>1,2</sup>, Camilla Meirelles Silva<sup>1,2</sup>, Juliana E. Toller-Kawahito<sup>1,2</sup>, Mikhael de Lima<sup>1,2</sup>, Daniele Carvalho Nascimento<sup>1,2</sup>, Ayda Henriques Schneider<sup>1,2</sup>, Italo J. Roberto Rosales<sup>1</sup>, David Calder<sup>1,2</sup>, Ronaldo Martins<sup>1,4</sup>, Italo J. Maria Nelson Benatti<sup>1</sup>, Leticia Pastorelli Bonjorno<sup>1</sup>, Marcela Ca Fernando Vilas<sup>1</sup>, Rodrigo Santana<sup>1</sup>, Valdes R. Bolidi<sup>1</sup>, Mari Antônio Pazin-Filho<sup>1</sup>, Luis Lambertini P. da Silva<sup>1,4</sup>, Larissa Doll Sabina Basati<sup>1</sup>, Alexandre Fabro<sup>1</sup>, Thais Masad<sup>1</sup>, Marisa Doll José Carlos Alves Filho<sup>1,2</sup>, Eunice Arruda<sup>1,4</sup>, Paulo Louzada Junior

Interdisciplinary Sciences: Computational Life Sciences (2021) 13:766–777  
<https://doi.org/10.1007/s12539-021-00462-3>

ORIGINAL RESEARCH ARTICLE

## HD5 and LL-37 Inhibit SARS-CoV and SARS-CoV-2 Binding to Human ACE2 by Molecular Simulation

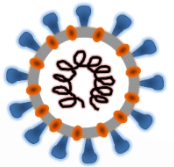
Daixi Li<sup>1</sup> · Peiqin Chen<sup>1</sup> · Ting Shi<sup>2</sup> · Aamir Mehmood<sup>2</sup> · Jingfei Qiu<sup>3</sup>



## Neutrophil extracellular traps contribute to immunothrombosis in COVID-19 acute respiratory distress syndrome

Elizabeth A Middleton<sup>1,2</sup>, Steven P Salvatore<sup>4,5</sup>, Loda<sup>4,5</sup>, Xue-Yan He<sup>3</sup>, Frederik Denorme<sup>1</sup>, Robert A Campbell<sup>1,2</sup>, David Ng<sup>3</sup>, ka<sup>4</sup>, Amelia Baxter-Stoltzfus<sup>4</sup>, Alain C Borczuk<sup>4,5</sup>, Massimo ith Manne<sup>1</sup>, Irina Portier<sup>1</sup>, Estelle S Harris<sup>2</sup>, Aaron C Petrey<sup>1</sup>, Anthony Iovino<sup>6,8</sup>, Lisa M Abegglen<sup>6,8</sup>, Andrew S Weyrich<sup>1</sup>, Egeblad<sup>3</sup>, Joshua D Schiffman<sup>1,6,8</sup>, Christian Con Yost<sup>1,6</sup>

Blood. 2020 Sep 3;136(10):1169–1179. doi: 10.1182/blood.2020007008.  
 DOI: 10.1182/blood.2020007008

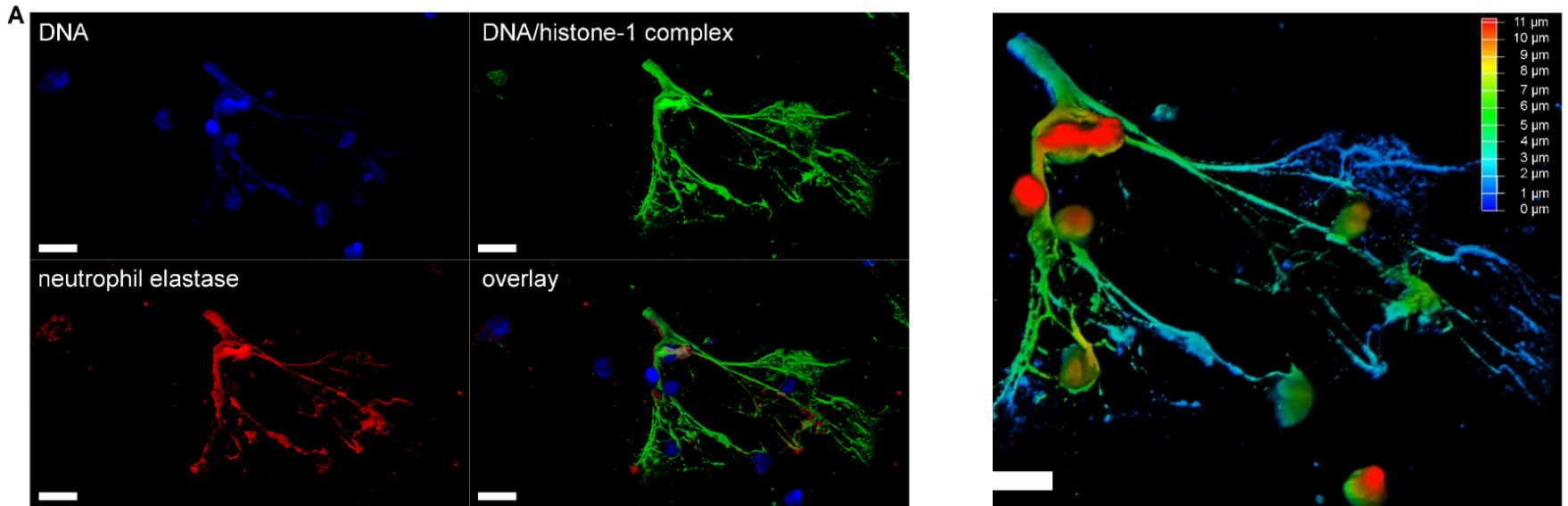


- Are NETs or AMPs counteracting SARS-CoV-2 and beneficial?
- Are NETs & AMPs detrimental in the COVID-19 pathogenesis?
- Is it beneficial for COVID-19 patients to modulate NETs & AMPs?



# NETs & AMPs interact with SARS-CoV-2

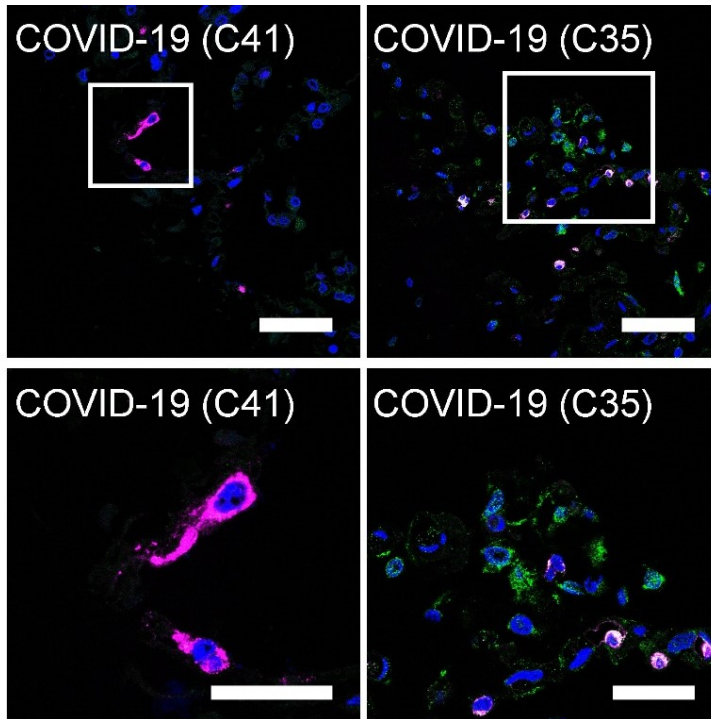
## NETs & SARS-CoV-2 *in vitro*, Timo Henneck



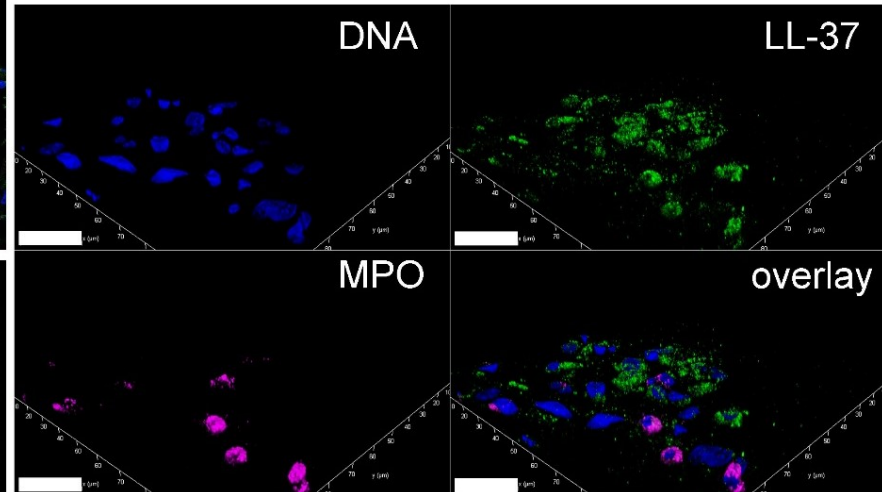
*In vitro*: Primary human blood-derived neutrophils release NETs in response to SARS-CoV-2.

# NETs & AMPs interact with SARS-CoV-2

## NETs & SARS-CoV-2 in human biopsies from COVID-19 patients



COVID-19 (C35) zoom 3D-image

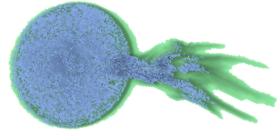


blue = DNA  
green = LL-37  
magenta =  
MPO/myeloperoxidase

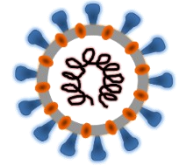
## Cationic antimicrobial peptide LL-37 stabilizes NETs

(Neumann et al., 2014; de Buhr et al., 2017, Meurer et al., 2020)

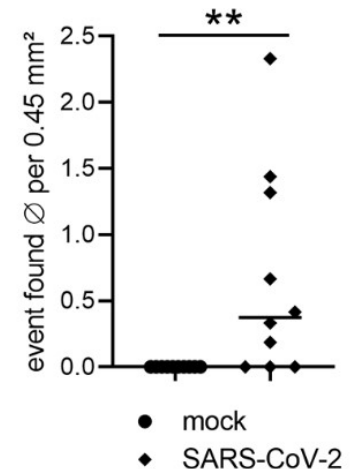
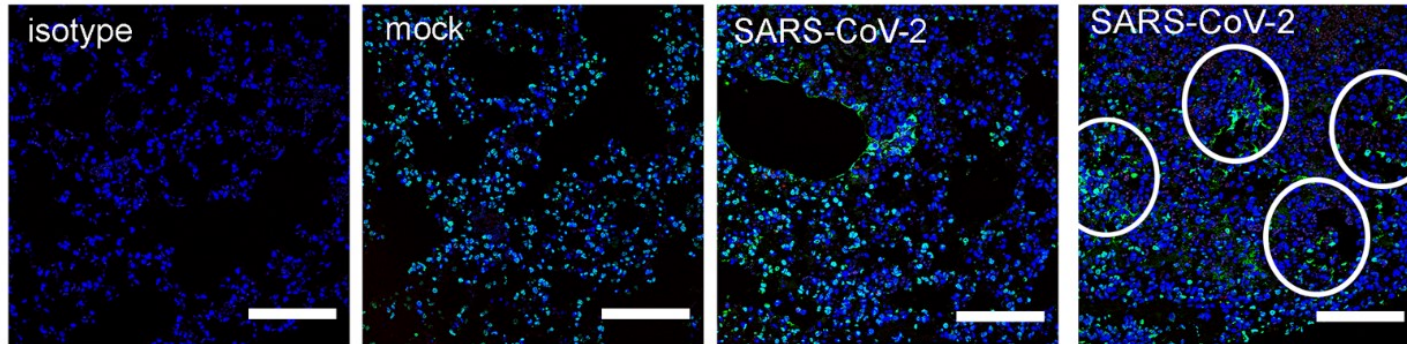
# NETs & AMPs interact with SARS-CoV-2



intranasally infected with  $10^5$  plaque forming units (p.f.u.) SARS-CoV-2 or mock



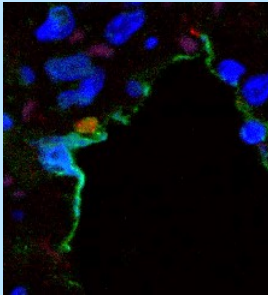
1, 3, 6 and 14 p.i. ten animals per group were euthanized and lungs were fixed in 4 % paraformaldehyde for microscopic examination.



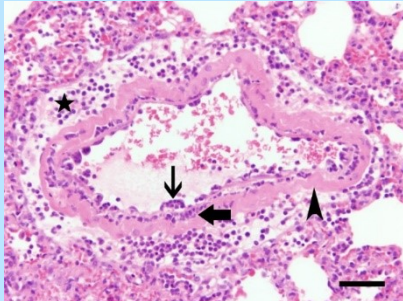
➤ Semiquantitative analysis of NET formation in SARS-CoV-2-infected animals was significantly increased compared to mock animals.

# NETs & AMPs interact with SARS-CoV-2

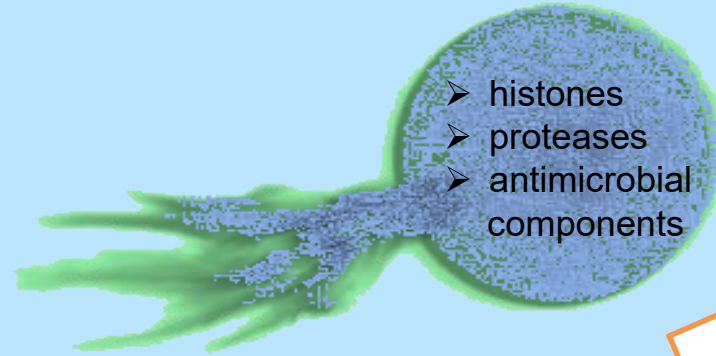
NETs are formed by infiltrating neutrophils into the lung as response to SARS-CoV-2-infections



Vasculitis during SARS-CoV-2 infection



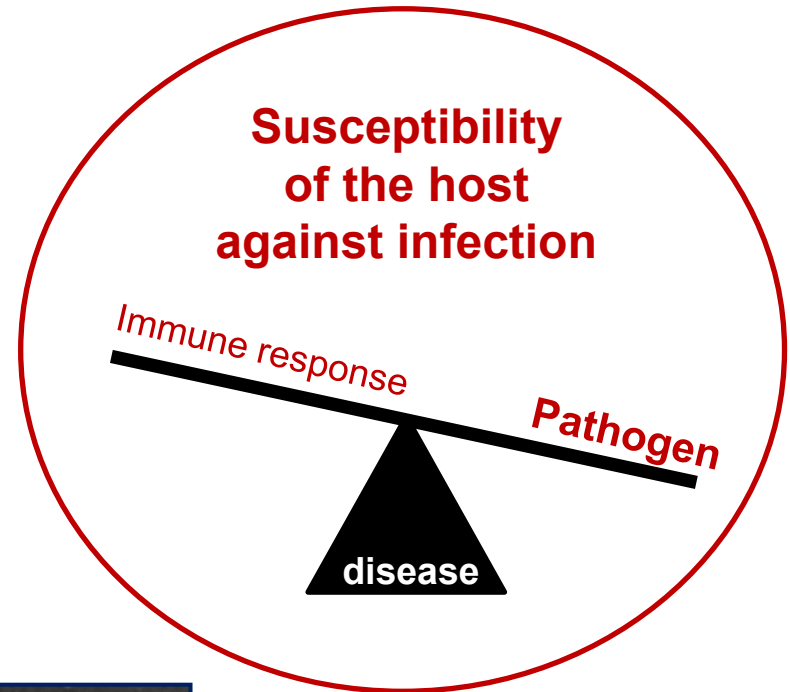
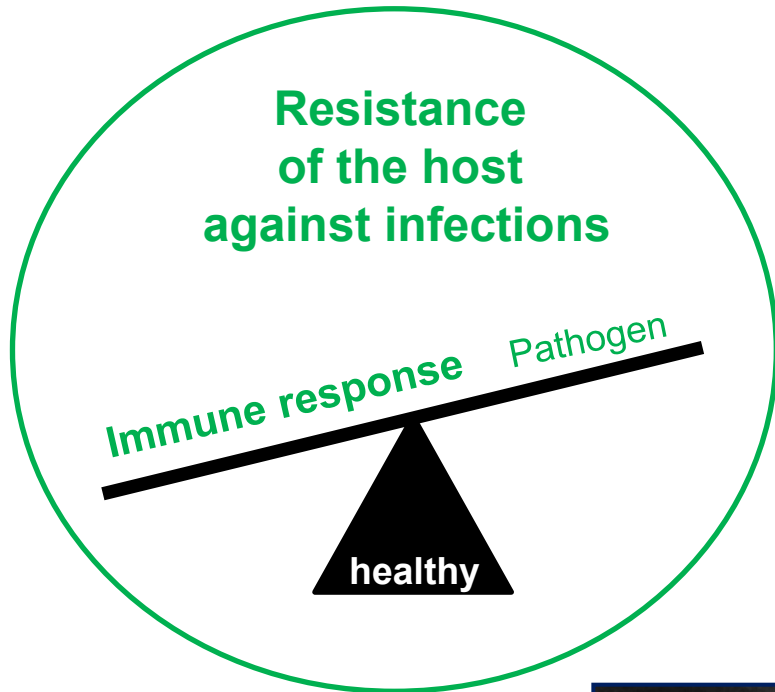
- Increased vascular permeability
- loss of barrier integrity, cytotoxicity
- microvascular injury)



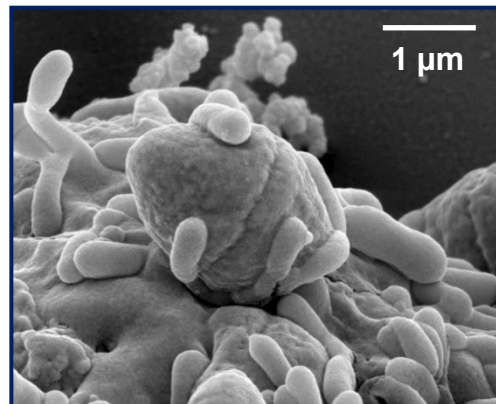
- histones
- proteases
- antimicrobial components

- pore-formation
- cytotoxicity (cationic charge and binding to phospholipid-membranes)

# Infections in human and animals



*Streptococcus pyogenes* entrapped by immune cell



von Köckritz-Blickwede et al., *Blood*, 2008

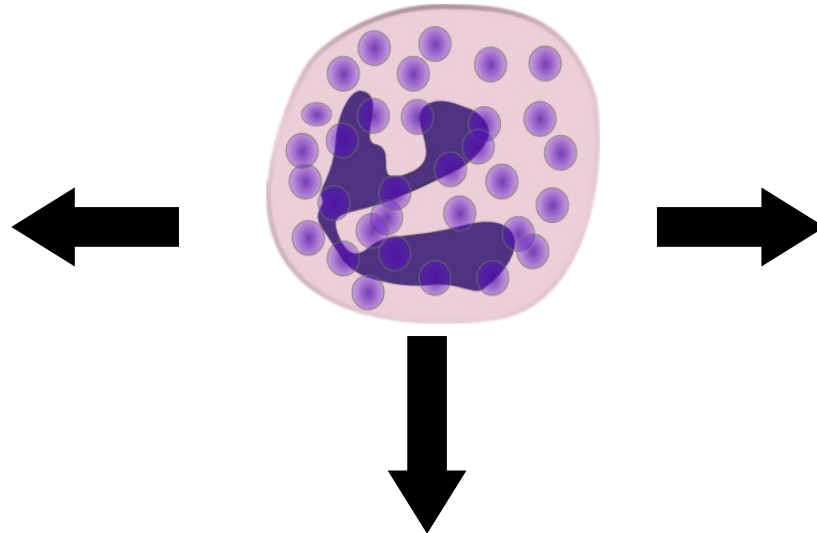
# Guardian cell of the immune system

## Neutrophil

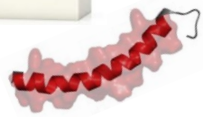
**Phagocytosis**  
(intracellular uptake and killing)



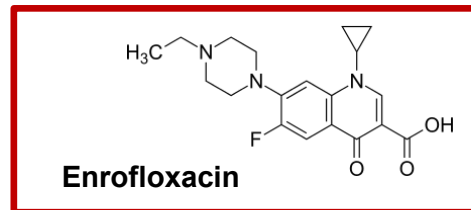
Guarea kunthiana



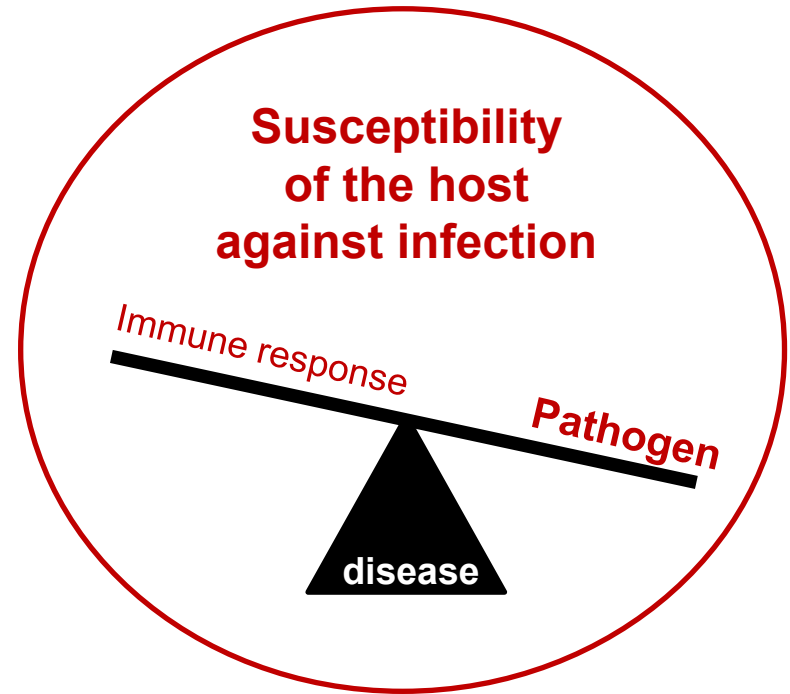
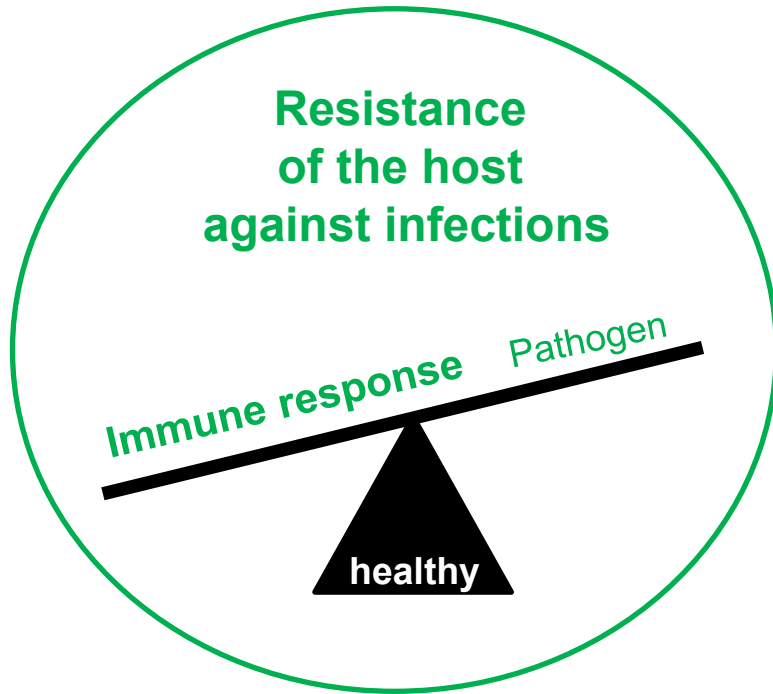
**Degranulation**  
(extracellular release of antimicrobial peptides and others)



**Neutrophil extracellular traps (NETs)**  
(extracellular entrapment and killing of bacteria)



# Infections in human and animals



Future task: Focus on the usage of antibiotics that modulate the innate immune response in a protective way?



# Infection Biochemistry

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**Research Group Infection Biochemistry**  
**Department of Biochemistry**  
**University of Veterinary Medicine Hannover, Germany**

# Support



Einzelantrag KO355214



Bundesministerium  
für Bildung  
und Forschung



und Forschung  
für Bildung  
Bundesministerium



Deutsche Akademie der  
Naturforscher Leopoldina



# DAAD

Deutscher Akademischer Austausch Dienst  
Servicio Alemán de Intercambio Académico



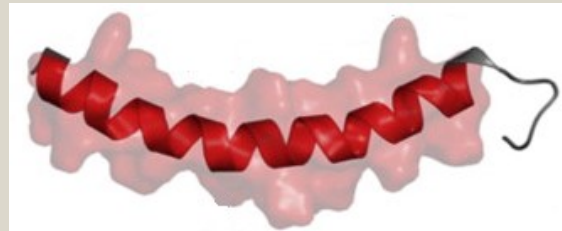
Akademie für Tiergesundheit



Lifelong  
Learning  
Programme

# Wie alte Waffen helfen, neue therapeutische Ansätze gegen Infektionskrankheiten zu finden

*„Old weapons against infectious diseases“*



Prof. Dr. Maren von Köckritz-Blickwede,  
Infection Biochemistry,  
Department of Biochemistry & Research Center for Emerging  
Infections and Zoonoses, TiHo Hannover, Germany

# Stärkung des Immunsystems

## Unser Ziel:

Suche nach neuen therapeutischen Ansätzen  
gegen Infektionskrankheiten:  
Naturprodukte, die das Immunsystem stärken!

Testung von Medizinalpflanzen  
aus dem tropischen  
Regenwald Costa Ricas:

- Verstärkung der Phagozytose  
von Neutrophilen
- Verstärkung der NETs

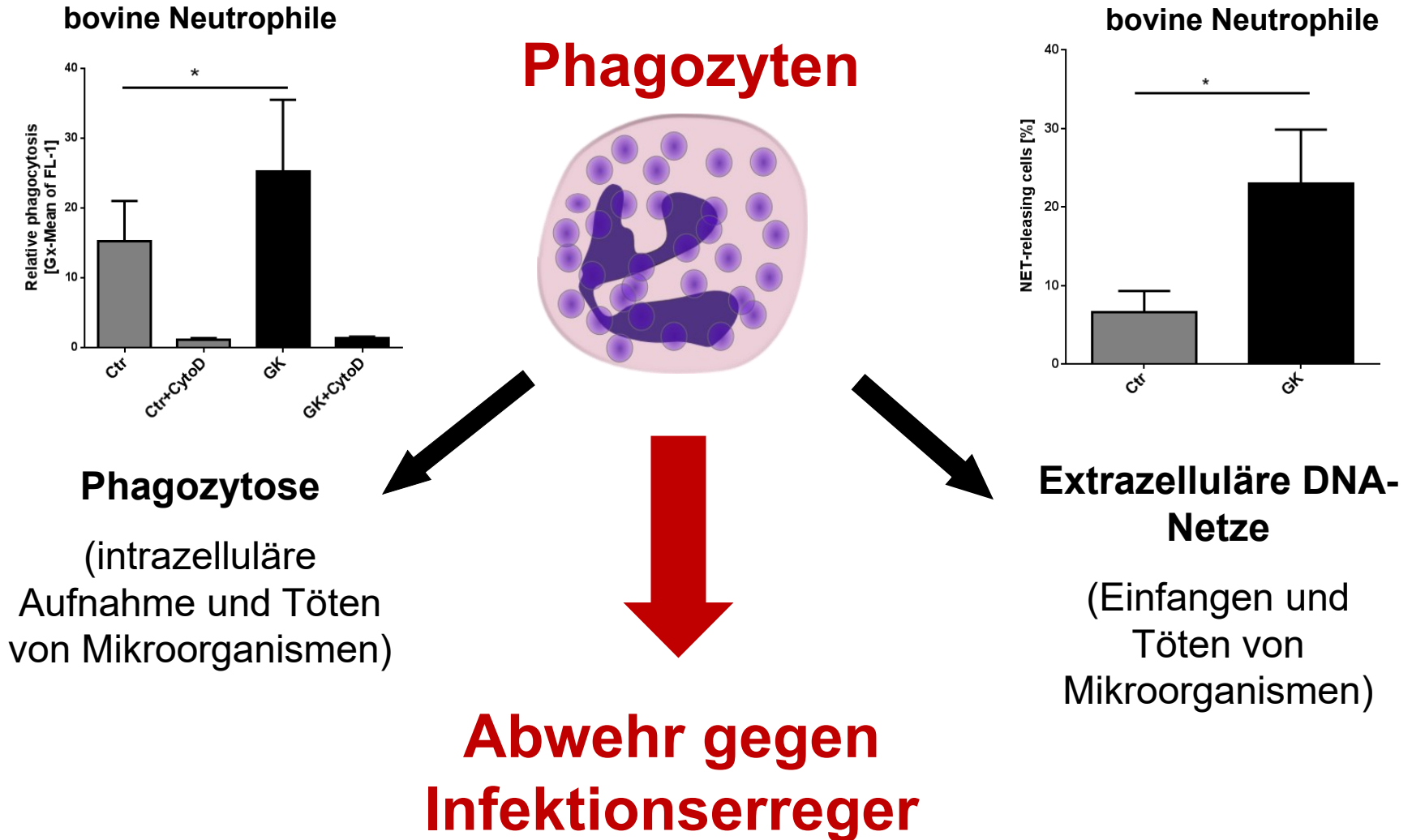


# GUKUBA stimuliert Immunzellen

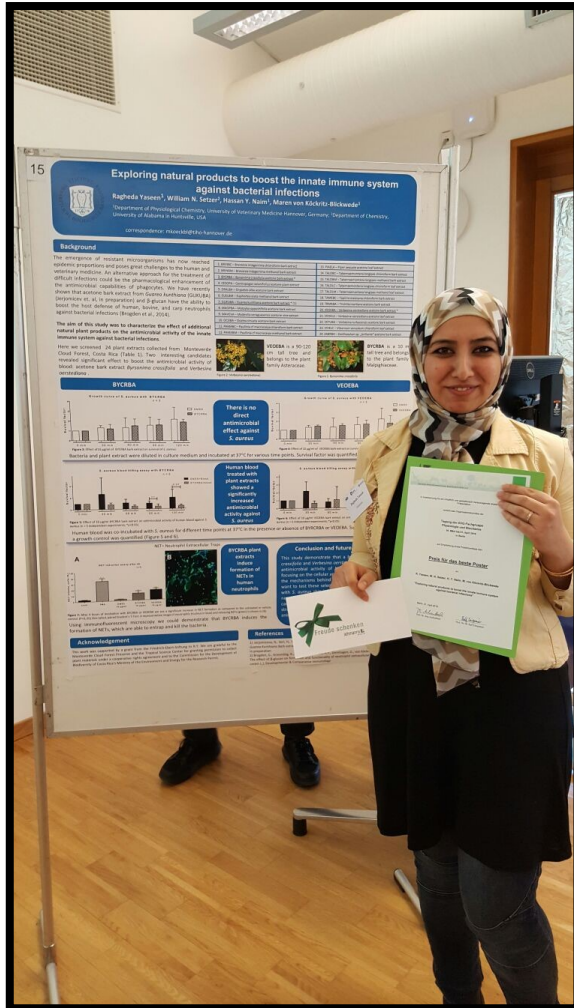
- Rindenextrakt von **Guarea kunthiana** (wächst im tropischen Bergwald von Süd- und Zentralamerika)
- Verwendung als Medizinalpflanze zur Behandlung von Depressionen und Asthma
- Rindensaft wird auch in Ecuador gegen Malaria eingesetzt.
- **Mechanismus unklar!!!**



# GUKUBA stimuliert Immunzellen



# Posterpreis für Ragheda Yaseen



*“Exploring natural products to boost the innate immune system against bacterial infections”*

Deutsche Veterinärmedizinische Gesellschaft  
Fachgruppe „Physiologie und Biochemie“,  
Berlin, April 2016



# Gum Arabic/Gummiarabikum

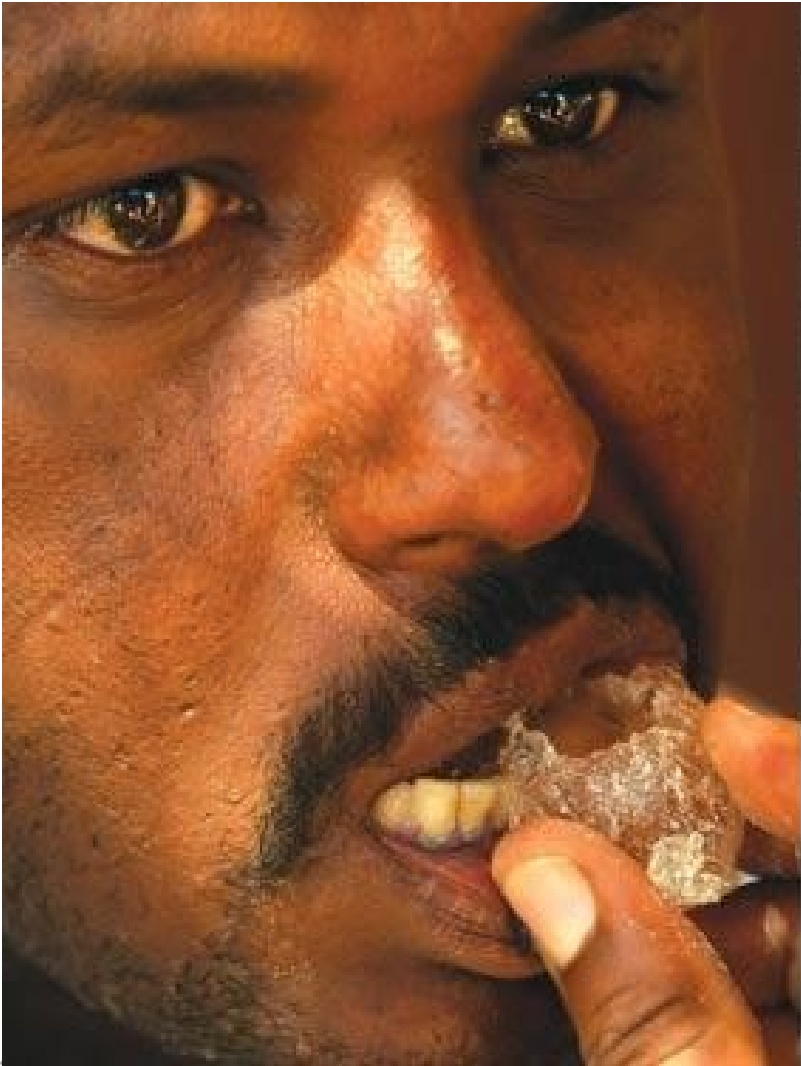
- ❖ Gum Arabic (GA), ist ein Exsudat aus dem Pflanzensaft von Verek-Akazien (*Acacia Senegal*) und Seyal-Akazien (*Vachellia seyal*)
- ❖ Zusammensetzung
  - Zucker
  - Glucuronsäure
  - Mineralien
  - Proteine



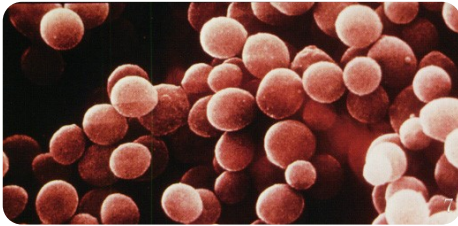
<http://www.fao.org/ag/agp/agpc/doc/Gallery/pictures/acasen/acasen1.jpg>



# Medizinalpflanze



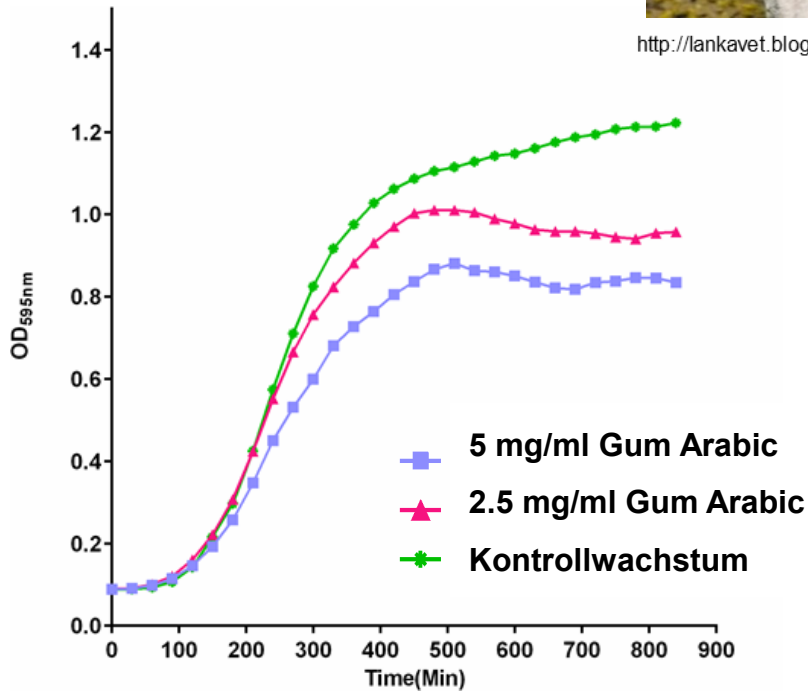
# Gum Arabic hemmt das Bakteriumwachstum



*Staphylococcus aureus*



<http://lankavet.blogspot.de/2013/01/mastitis-in-cows.html>



Shima Baein

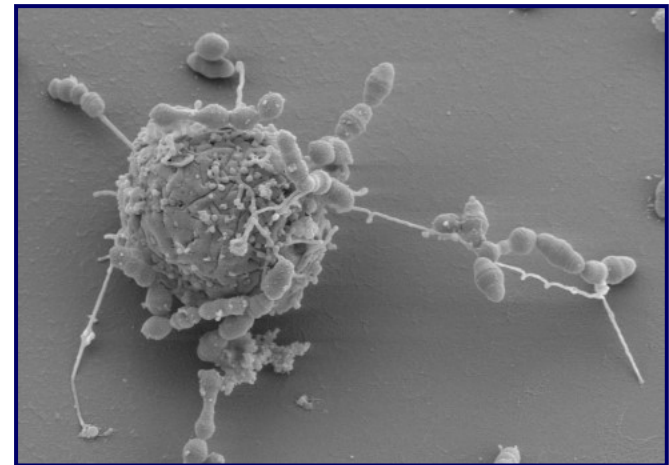


Nicole de Buhr

# Institut für Physiologische Chemie

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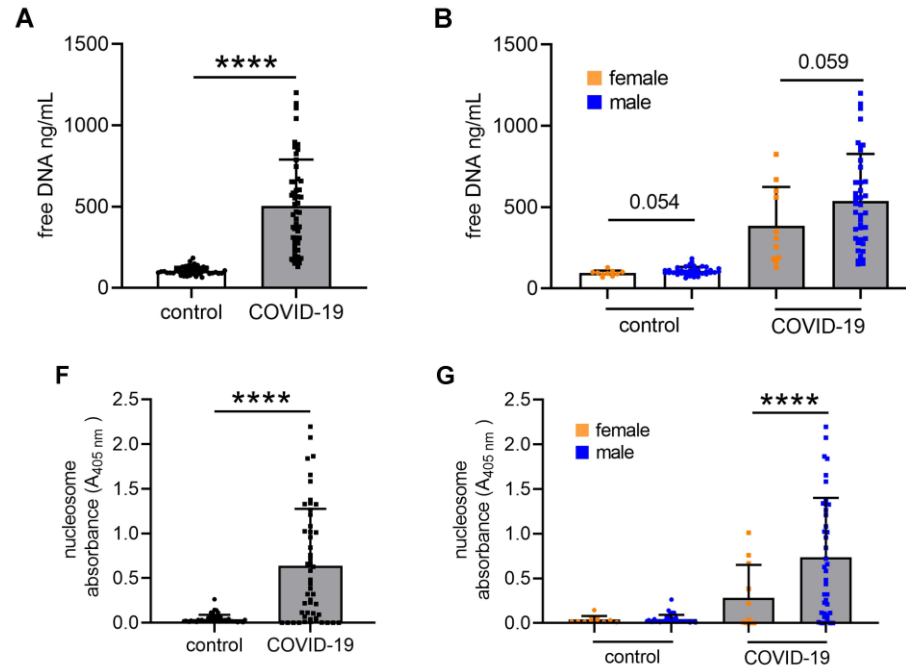
**Unser Ziel:**  
**Suche nach neuen therapeutischen Ansätzen  
gegen Infektionskrankheiten:  
Stärkung des Immunsystems!**



Fotos: Abwehrzellen fangen Bakterien mit NETzen.

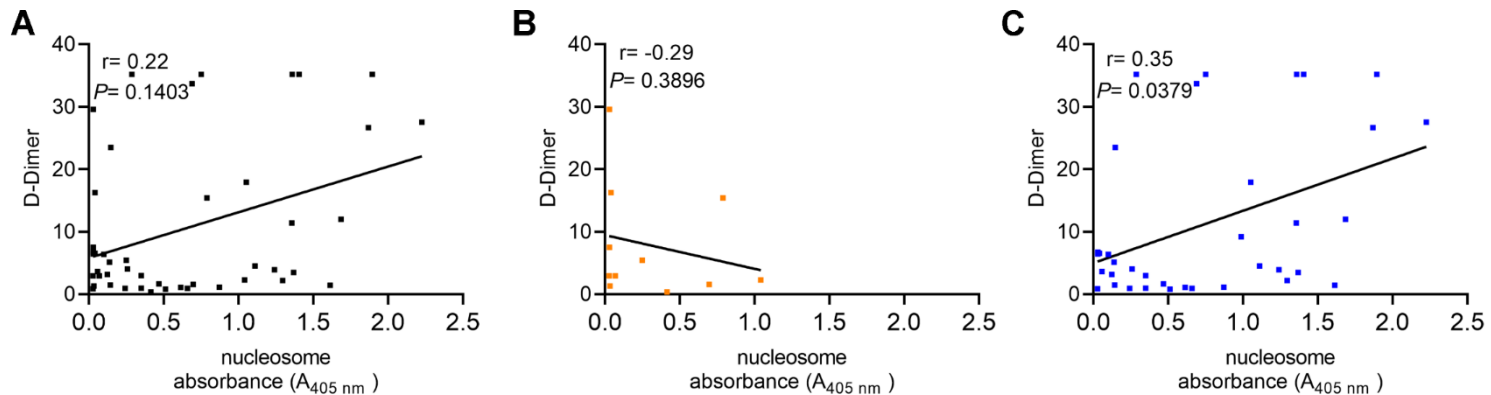
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## NETs & SARS-CoV-2 in serum of COVID-19 patients



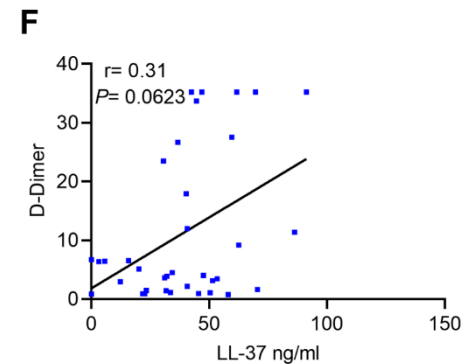
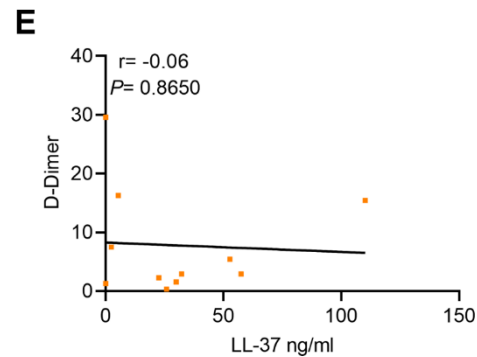
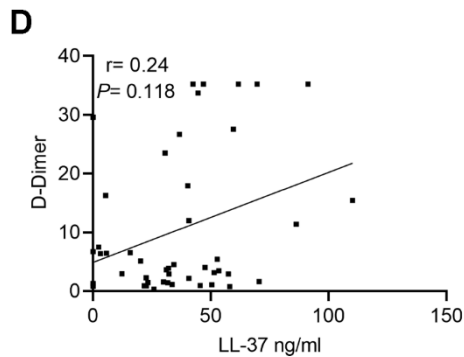
Increased free eDNA and nucleosome level as NET marker were detected in COVID-19 patients compared to healthy controls.

## NETs & SARS-CoV-2 in serum of COVID-19 patients



- Severe infection of COVID-19 exhibit more obvious dysregulated coagulation function compared with mild cases. This is shown by a higher level of D-dimer (Snawa et al., 2021)
- Nucleosomes as marker for NETs strongly correlated with D-dimer
- This confirms previous published data that NETs and fibrin from a composite network within thrombin that triggers the risk for thrombosis in COVID-patients.

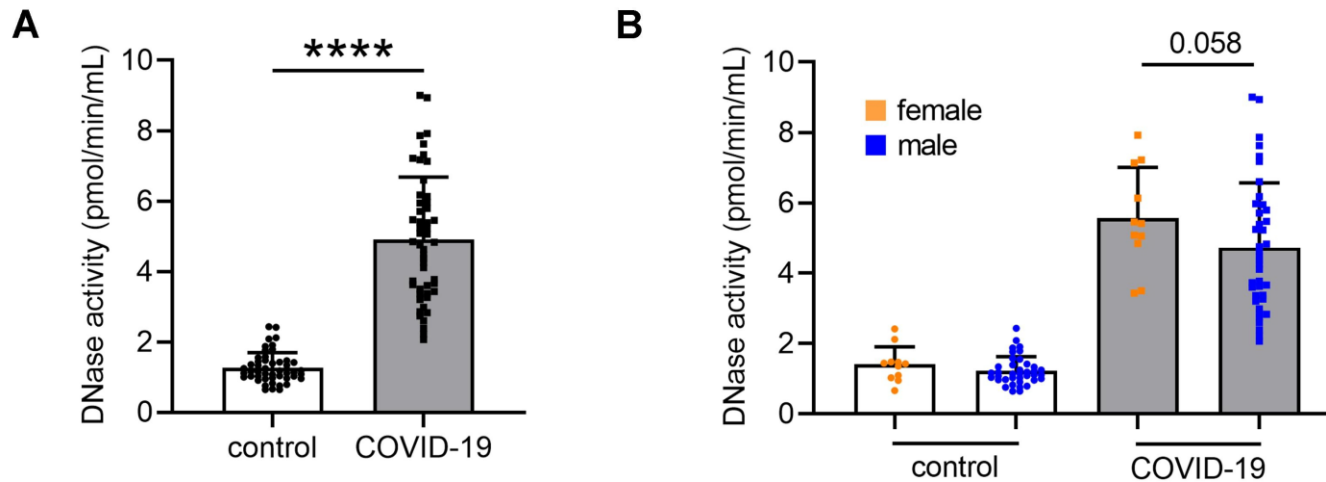
## NETs & SARS-CoV-2 in serum of COVID-19 patients



**Cationic antimicrobial peptide LL-37, stabilizes NETs**

- **LL-37 level correlate with D-Dimer level in male COVID-patients**
- **Potential additional role of LL-37 in the risk of NET-associated thrombosis of male COVID-patients by stabilizing NETs?**

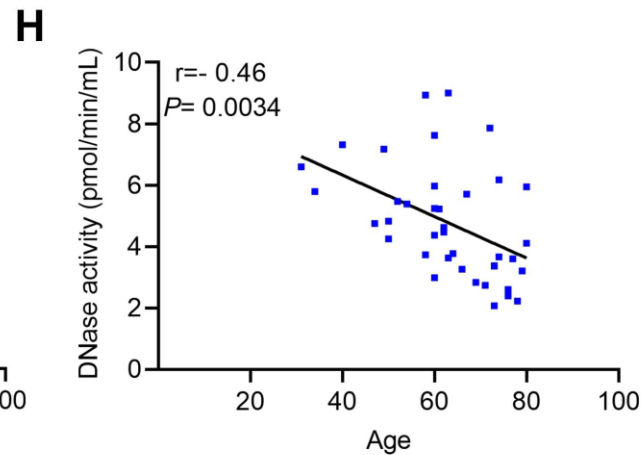
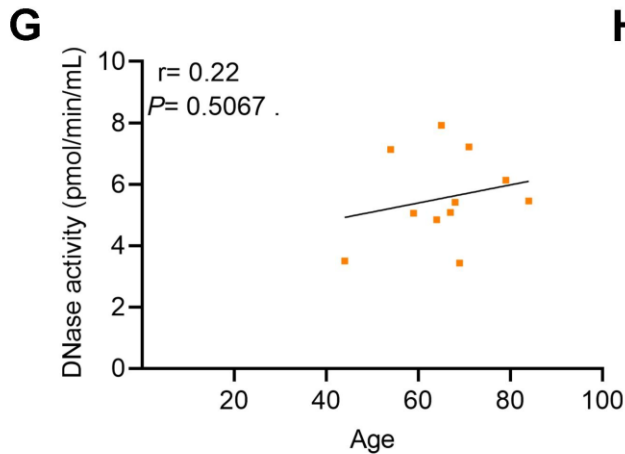
## NETs degradation by nucleases



Host  
DNases  
to  
degrade  
NETs

- Increased DNase activity as a response to NET release in COVID patients compared to healthy control.
- No difference in male compared to female.

## NET degradation by nucleases



Host  
DNases  
to  
degrade  
NETs

- Increased DNase activity as a response to NET release in COVID patients compared to healthy control.
- No difference in male compared to female.
- **Lower DNase activity in elderly male patients.**



## Summary: NETs & SARS-CoV-2

1. NETs are highly discussed in literature to contribute to COVID-19-severity.
2. Our data confirm increased NET marker in COVID-19 patients compared to healthy controls.
3. NETs are confirmed in biopsies from COVID-19 patients.
4. NET-stabilizing peptide LL-37 is shown to bind NETs that are produced by neutrophils in COVID-patients.
5. LL-37 level correlate with D-Dimer level similar as NET-marker in male COVID-patients, indicating a potential additional role of LL-37 in the risk of NET-associated thrombosis of male COVID-patients.
6. LL-37 in NETs is confirmed in biopsies from COVID-19 patients.
7. Increased DNase activity is confirmed in COVID-patients, showing that patients react to increased NET-formation by producing NET-degrading enzymes e.g. DNase 1.
8. Male elderly patients show decreased DNase 1 activity, hypothesizing a potential risk factor for those patients in their ability to degrade NETs.

→ **Two risk factors of male elderly patients: Reduced nuclease activity and increased LL-37 level which lead to inefficient NET degradation and**

## NETs & SARS-CoV-2

### *Research Article*

### *Impaired degradation of neutrophil extracellular traps: a possible severity-factor of elderly male COVID-19-patients*

Nicole de Buhr<sup>a,b\*</sup>, Ann Parplys<sup>c\*</sup>, Maria Schröder<sup>d\*</sup>, Timo Henneck<sup>a,b</sup>, Berfin Schaumburg<sup>c</sup>, Dominik Jarczak<sup>d</sup>, Axel Nierhaus<sup>d</sup>, Stefan Kluge<sup>d</sup>, Karin Klingel<sup>e</sup>, Gülsah Gabriel<sup>c,ff</sup>, Maren von Köckritz-Blickwede<sup>a,b,#</sup>

*Major revision, Journal of Innate Immunity*

